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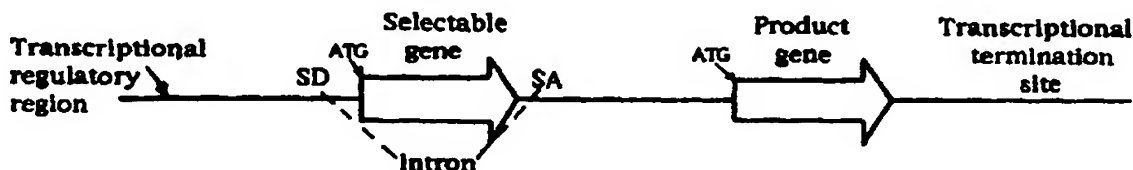
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(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS



(57) Abstract

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing host cells, a method of producing a protein of interest in high yields and a method of producing eukaryotic cells having multiple copies of a sequence encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells in a functional form has provided the key to understanding many fundamental biological processes, and has made possible the production of important proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several common problems exist that may limit the efficiency with which a gene encoding a desired protein can be introduced into and expressed in a host cell. One problem is knowing when the gene has been successfully transferred into recipient cells. A second problem is distinguishing between those cells that contain the gene and those that have survived the transfer procedures but do not contain the gene. A third problem is identifying and isolating those cells that contain the gene and that are expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic cells tend to be highly inefficient. Of the cells in a given culture, only a small proportion take up and express exogenously added DNA, and an even smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene encoding a desired protein typically is achieved by introducing into the same cells another gene, commonly referred to as a selectable gene, that encodes a selectable marker. A selectable marker is a protein that is necessary for the growth or survival of a host cell under the particular culture conditions chosen, such as an enzyme that confers resistance to an antibiotic or other drug, or an enzyme that compensates for a metabolic or catabolic defect in the host cell. For example, selectable genes commonly used with eukaryotic cells include the genes for aminoglycoside phosphotransferase (APH), hygromycin phosphotransferase (hyg), dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin, glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene on the basis of expression by the host cell of a second incorporated gene encoding a selectable marker is referred to as cotransfection (or cotransfection). In that method, a gene encoding a desired polypeptide and a selection gene typically are introduced into the host cell simultaneously, although they may be introduced sequentially. In the case of simultaneous cotransfection, the gene encoding the desired polypeptide

and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by
5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency
10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter
15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and
20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

Enhancers stimulate transcription from a linked promoter. Unlike
25 promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters
30 listed above also contain strong enhancers. Bendig, Genetic Engineering, 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing the gene copy number in the host cell. One method for obtaining high gene copy number is to directly introduce into the host cell multiple copies of
35 the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransfection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high copy number. Furthermore, because no generally applicable, convenient
40 method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning
45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors

derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective
10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under
15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell is cotransfected with a product gene encoding a desired protein and a DHFR
20 gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Nat. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are
25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic
30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-
35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant DHFR gene that encodes a protein with reduced sensitivity to methotrexate
40 may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a *neo^r* gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such recombinant host cells in a single step. One strategy involves co-transfecting host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent
5 appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621
10 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320
[1988]; Jang et al., J. Virol., 63:1651 [1989]).

15 A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected
20 with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that
25 the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

30 Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This
35 vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by
40 expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the
45 numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10 SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates expression of the product gene using the transcriptional regulatory region.

20 In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

25 In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

30 After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence (Δ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 3 used for expression of anti-IgE V_H.

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron-(WT ras SD)-tPA expression vector of Example 1.

Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [Δ GT]) and from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the Δ GT pool) were exposed to 200nM Mtx to select for DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron-(WT ras SD)-TNFr-IgG expression vector of Example 2.

Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12.CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

Figure 8 depicts schematically the DNA construct used for expression of the V_L of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V_H expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V_L expression vector of Example 3.

5 Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V_H) and light (V_L) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

10 DESCRIPTION OF THE PREFERRED EMBODIMENTS

Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the
15 chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of
20 growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in
25 Enzymology, 185: 537-566 (1990), for a review of these.

TABLE 1
Selectable Genes and their Selection Agents

Selection Agent	Selectable Gene
Methotrexate	Dihydrofolate reductase
30 Cadmium	Metallothionein
PALA	CAD
Xyl-A-or adenosine and 2'-deoxycoformycin	Adenosine deaminase
35 Adenine, azaserine, and coformycin	Adenylate deaminase
6-Azaauridine, pyrazofuran	UMP Synthetase
Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs <i>e.g.</i> adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	β -Aspartyl hydroxamate or Albizziin	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	α -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na ⁺ K ⁺ -ATPase

The preferred selectable gene is an amplifiable gene. As used herein, the term "amplifiable gene" refers to a gene which is amplified (*i.e.* additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (*i.e.* an amplifiable marker) which is required for growth of eukaryotic cells under those conditions. For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mtx. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko *et al.*, *supra*).

As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary nutrient solutions. In addition, any of the media described in Ham and Wallace, Meth. Enz., 58:44 (1979), Barnes and Sato, Anal. Biochem., 102:255

(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be
5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as GentamycinTM drug), trace elements (defined as inorganic compounds usually
10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with
15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table
20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA
25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a
30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a cis-acting DNA element,
35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the
40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product
45 genes suitably encode a peptide, or may encode a polypeptide sequence of

amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and β -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- β ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- β 1, TGF- β 2, TGF- β 3, TGF- β 4, or TGF- β 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- β , TGF- α , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- γ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or *de novo* synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gattermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer to the *in vitro* amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice acceptor site occurs within the consensus sequence (U/C)₁₁NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability and/or copy number of a transcribed sequence in order to allow for elevated levels of a RNA sequence of interest. In general, the methods of the present invention involve transfecting a eukaryotic host cell with an expression vector comprising both a product gene encoding a desired polypeptide and a selectable gene (preferably an amplifiable gene).

Selectable genes and product genes may be obtained from genomic DNA, cDNA transcribed from cellular RNA, or by *in vitro* synthesis. For example, libraries are screened with probes (such as antibodies or oligonucleotides of about 20-80 bases) designed to identify the selectable gene or the product gene (or the protein(s) encoded thereby). Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the selectable gene or product gene is to use PCR methodology as described in section 14 of Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues known to contain the selectable gene or product gene. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use ³²P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is preceded by DNA encoding a signal sequence having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the expression vector, or it may be a part of the selectable gene or product gene that is inserted into the expression vector. If a heterologous signal sequence is used, it preferably is one that is recognized and processed (*i.e.*, cleaved by a signal peptidase) by the host cell. For yeast secretion the native signal sequence may be substituted by, *e.g.*, the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α -factor leaders, the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression the native signal sequence

of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spliced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfectants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as *de novo* synthesis or *in vitro* deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general
5 proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction
10 endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single
15 transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem., 255:2073 [1980]) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Req., 7:149 [1968]; and Holland,
20 Biochemistry, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the
25 additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and
30 promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30
35 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as
40 polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters,
45 and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a HindIII E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; , Reyes et al., Nature, 297:598-601 (1982) on expression of human β -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon β 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA

fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 μ plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvencourt et al., J. Bacteriol., 737 (1983)], *Yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*

(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8
5 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda*
10 cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain
15 the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen.,
20 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

25 However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL
30 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dp12.CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells
35 (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse
40 mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient
45 media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly ³²P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescens, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay
5 of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels,
10 fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is
15 measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human
20 origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged
25 to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate
30 precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and
35 literature references cited herein are expressly incorporated by reference.

EXAMPLE 1

tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such
40 as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the
45 cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V_H) gene (Eaton et al., Biochem., 25:8343 [1986]).

5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that
10 comprises the splice donor site. Those sequences were altered by overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59
15 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice
20 site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control (Δ GT). A polylinker was
25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site (Δ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease
35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 μ l 1/10 Tris EDTA. Then, 10 μ g of DNA was incubated with 10⁷ CHO.dp12 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before
40 electroporation at 400 volts and 330 μ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

45 To prepare the mRNA, RNA was extracted from 5 x 10⁷ cells which were grown from pools of more than 200 clones derived from the stable

transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10µg of mRNA was then subjected to Northern blotting which involved running the mRNA on
5 a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and
10 spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only
15 full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after duplicate electroporations with the three intron vectors described above and from a conventional vector that has a CMV promoter driving tPA and a
20 SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector efficiently generates colonies and does not vary significantly from the ΔGT
25 construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluency. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was
30 coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling
35 the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open
40 reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors
45 (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA, SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing (Δ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2 μ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR^r cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

EXAMPLE 2

TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., Proc. Natl. Acad. Sci. USA, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site (Δ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 $\mu\text{g/ml}$ when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, contrary to expectations, it was discovered that isolation of high product expressing host DHFR⁺ cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

EXAMPLE 3

Antibody production using a dicistronic expression vector

The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the promoter/ enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNF α -IgG constructs. Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression. All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1 μM and achieved a greater than 2 fold increase in antibody titer. The 1 μM Mtx resistant pool achieved a titer of 41 mg/L when grown under optimal conditions in suspension culture.

The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S^{35} cysteine and methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50 μ Ci each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain expressed by the pool.

CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNFr immunoadhesin or anti-IgE V_H .

The homogeneity of expression of clones generated by the ras splice donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived from the conventional vector may be the result of breaks in the tPA or

TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or
5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased
10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely
15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain
25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain
30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 μ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly
45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 10 (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
 (A) ADDRESSEE: Genentech, Inc.
 (B) STREET: 460 Point San Bruno Blvd
 (C) CITY: South San Francisco
 15 (D) STATE: California
 (E) COUNTRY: USA
 (F) ZIP: 94080
- (v) COMPUTER READABLE FORM:
 20 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
 (B) COMPUTER: IBM PC compatible
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 (D) SOFTWARE: patin (Genentech)
- 25 (vi) CURRENT APPLICATION DATA:
 (A) APPLICATION NUMBER:
 (B) FILING DATE:
 (C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
 (A) APPLICATION NUMBER: 08/286740
 (B) FILING DATE: 05-AUG-1994
- (viii) ATTORNEY/AGENT INFORMATION:
 35 (A) NAME: Lee, Wendy M.
 (B) REGISTRATION NUMBER: 00,000
 (C) REFERENCE/DOCKET NUMBER: 798PCT
- (ix) TELECOMMUNICATION INFORMATION:
 40 (A) TELEPHONE: 415/225-1994
 (B) TELEFAX: 415/952-9881
 (C) TELEX: 910/371-7168

(2) INFORMATION FOR SEQ ID NO:1:

- 45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 7360 bases
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 50 (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50

TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100

60 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150

ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200

65 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250

ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300

5 AAATGGCCCCG CCTGGCATT A TGCCAGTAC ATGACCTTAT GGGACTTTCC 350

TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400

10 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450

TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500

15 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550

AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

20 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

25 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

30 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

35 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

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55 GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

60 ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300

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65 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATATA 1400

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5 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGCTACTCCA 1550

10 GGTCAACTGC ACCTCGGTTC TAAGCTTGGG CTGCAGGTCG CCGTGAATTT 1600

AAGGGACGCT GTGAAGCAAT CATGGATGCA ATGAAGAGAG GGCTCTGCTG 1650

15 TGTGCTGCTG CTGTGTGGAG CAGTCTTCGT TTCGCCCAGC CAGGAAATCC 1700

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25 TGTGAGCTCT CCGGCTACGG CAAGCATGAG GCCTTGTCTC CTTTCTATTC 3000

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30 CATCACAACA TTTACTTAAC AGAACAGTCA CCGACAACAT GCTGTGTGCT 3100

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50 TCTCCAGACC CACCACACCG CAGAAGCGGG ACGAGACCCT ACAGGAGAGG 3450

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60 GCCTCTCCAG GAATGCCTCC TCCCTGGGCA GAAGTGGGGG GAATTCAATC 3600

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35 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC 5500

CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 5550

GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT 5600

45 ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC 5650

ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC TGCCATAACC 5700

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55 GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT GTAAC TCGCC 5800

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60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900

TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG 5950

65 AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT TCCGGCTGGC 6000

TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050

5 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100

ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150

10 GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA 6200

CTCATATATA CTTTAGATTG ATTTAAAACT TCATTTTAA TTTAAAAGGA 6250

15 TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAAT CCCTTAACGT 6300

GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350

20 TTCTTGAGAT CCTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA 6400

25 AACCACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT 6450

CTTTTTCCGA AGGTAAGTGG CTTGAGCAGA GCGCAGATAC CAAATACTGT 6500

30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550

CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT 6600

35 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650

40 TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA CAGCCCAGCT 6700

TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750

45 GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800

CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGGAAACG 6850

50 CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT 6900

55 CGATTTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG 6950

CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA 7000

60 TGTTCTTTCC TGCGTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC 7050

TTTGAGTGAG CTGATACCGC TCGCCGAGC CGAACGACCG AGCGCAGCGA 7100

65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCAT TAATCCAGCT GGCACGACAG GTTTCCTGAC 7200
5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250
TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300
10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350
TACGAATTAA 7360

15

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
20 (A) LENGTH: 6889 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
30 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTCCGC GTTACATAAC 100
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
35 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
45 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
50 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
60 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600
TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650
65 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950

15 CCTGGTTCTC CATTCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000

ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050

20 TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100

25 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150

GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

30 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

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35 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350

40 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTTATAA 1400

GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTGGCTTC GTTAGAACGC 1450

45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

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50 GGTCAACTGC ACCTCGGTTC TATCGATTGA ATTCCCCGGC CATAGCTGTC 1600

55 TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650

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60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750

TATCCACCCT CAAAATAATT CGATTTGCTG TACCAAGTGC CACAAAGGAA 1800

65 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGAAGTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG 1900
5 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950
CTTGACACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000
10 CGGCATTATT GGAGTGAAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG 2050
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAAACAG AACACCGTGT 2100
15 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150
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20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250
25 AGCTCAAAAC CCCACTTGGT GACACAAC TC ACACATGCCC ACGGTGCCCA 2300
GAGCCCAAAT CTTGTGACAC ACCTCCCCCG TGCCACGGT GCCCAGAGCC 2350
30 CAAATCTTGT GACACACCTC CCCCATGCCC ACGGTGCCCA GAGCCCAAAT 2400
CTTGTGACAC ACCTCCCCCA TGCCACGGT GCCCAGCACC TGAATCCTG 2450
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCA AAACCAAGG ATACCCTTAT 2500
40 GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550
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45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650
GGTCAGCGTC CTCACCGTCC TGCACCAGGA CTGGCTGAAC GGCAAGGAGT 2700
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAAACC 2750
55 ATCTCCAAAA CCAAAGGACA GCCCCGAGAA CCACAGGTGT ACACCCTGCC 2800
CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTCAGCCTG ACCTGCCTGG 2850
60 TCAAAGGCTT CTACCCAGC GACATCGCCG TGGAGTGGGA GAGCAGCGGG 2900
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950
65 CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC 3000

AGGGGAACAT CTTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC 3050

5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAATGAG TCGACGGCC 3100

GGGGATCCTC TAGAGTCGAC CTGCAGAAGC TTGGCCGCCA TGGCCCAACT 3150

10 TGTTTATTGC AGCTTATAAT GGTTACAAAT AAAGCAATAG CATCACAAAT 3200

TTCACAAATA AAGCATTTTT TCACTGCAT TCTAGTTGTG GTTGTCCAA 3250

15 ACTCATCAAT GSTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300

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20 CTTCTGAGGC GGAAAGAACC AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400

GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450

AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500

30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550

TAACTCCGCC CATCCCGCCC CTAAGTCCGC CCAGTTCCGC CCATTCTCCG 3600

35 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC 3650

GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG 3700

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45 GTCGTGACTG GGAAAACCCT GGC GTTACCC AACTTAATCG CCTTGCAGCA 3800

CATCCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCCC GCACCGATCG 3850

50 CCCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900

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AACCATAGTA CGCGCCCTGT AGCGGCGCAT TAAGCGCGGC GGGTGTGGTG 4000

60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050

TTTCGCTTTC TTCCCTTCCT TTCTCGCCAC GTTCGCCGC TTTCCCGTC 4100

65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAAACT TGATTGGGT GATGGTTCAC GTAGTGGGCC 4200

5 ATCGCCCTGA TAGACGGTTT TCGCCCTTT GACGTTGGAG TCCACGTTCT 4250

TTAATAGTGG ACTCTTGTTT CAACTGGAA CAACACTCAA CCCTATCTCG 4300

10 GGCTATTCTT TTGATTATA AGGGATTTG CCGATTTCGG CCTATTGGTT 4350

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15 TAACGTTTAC AATTTTATGG TGCACCTCTCA GTACAATCTG CTCTGATGCC 4450

GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500

20 CCCCACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550

25 CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600

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30 AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT 4700

AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG 4750

35 GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC 4800

40 ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG 4850

TATGAGTATT CAACATTTCC GTGTCGCCCT TATTCCCTTT TTTGCGGCAT 4900

45 TTTGCCTTCC TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT 4950

GCTGAAGATC AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA 5000

50 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA 5050

55 TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100

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60 GGTTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG 5200

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65 AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT 5300

GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350
TGAATGAAGC CATAACAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400
5 ATGGCAACAA CGTTGCGCAA ACTATTAACT GGCGAACTAC TTACTCTAGC 5450
TTCCCGGCAA CAATTAATAG ACTGGATGGA GGCGGATAAA GTTGCAGGAC 5500
CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGTTTATTGC TGATAAATCT 5550
15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600
TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA 5650
20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700
AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA 5750
TTTAAAACTT CATTTTAAAT TTAAAAGGAT CTAGGTGAAG ATCCTTTTTG 5800
30 ATAATCTCAT GACCAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG 5850
TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT 5900
35 GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG 5950
TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC 6000
TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT 6050
45 AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100
TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150
50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTCGGGCTG 6200
AACGGGGGGT TCGTGCACAC AGCCCAGCTT GGAGCGAACG ACCTACACCG 6250
AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA 6300
60 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA 6350
GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400
65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTTGTG ATGCTCGTCA 6450

GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACGCGGCCT TTTTACGGTT 6500
5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT GCGTTATCCC 6550
CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATAACCGCT 6600
10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650
AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTCAAT 6700
15 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC 6750
20 AACGCAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA 6800
CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT 6850
25 TTCACACAGG AAACAGCTAT GACCATGATT ACGAATTAA 6889

30 (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6557 bases
(B) TYPE: nucleic acid
35 (C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50
GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100
45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150
TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA 200
50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250
55 CGCCCAGTTC CGCCCATTCT CCGCCCCATG GCTGACTAAT TTTTTTTATT 300
TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG 350
60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAAGCTA GCTTATCCGG 400
CCGGGAACGG TGCATTGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450
65 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500

TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA 550
5 AGAACGGAGA CCTACCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600
CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAAACAGA ATCTGGTGAT 650
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750
15 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA 800
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTCGGAGGCA 850
20 GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT 900
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950
TGATTTGGGG AAATATAAAC CTCTCCCAGA ATACCAGGC GTCCTCTCTG 1000
30 AGGTCCAGGA GGAAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100
35 ATGCATTTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCCTTGG 1150
40 CTTCGTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC 1200
ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGGTTCTAT CGATTGAATT 1300
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350
50 GGAGTACATT CAGAAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA 1400
55 GCCAGGGGGC TCACTCCGTT TGTCTGTGC AGTTTCTGGC TACTCCATCA 1450
CCTCCGGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500
60 GAATGGGTTG CATCGATTAC GTATGCCGGA TCGACTAACT ATAACCCTAG 1550
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCGT CTATTATTGT 1650

GCTCGAGGCA GCCACTATTT CGGCGCCTGG CACTTCGCCG TGTGGGGTCA 1700
5 AGGAACCCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750
TCCCCCTGGC ACCCTCCTCC AAGAGCACCT CTGGGGGCAC AGCGGCCCTG 1800
10 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCTGGAA 1850
CTCAGGCGCC CTGACCAGCG GCGTGACAC CTTCCCGGCT GTCCTACAGT 1900
15 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGA CTGTGCC CTCTAGCAGC 1950
TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGCAACAC 2000
20 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050
GCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCCTC 2100
TTCCCCCAA AACCAAGGA CACCCTCATG ATCTCCCGGA CCCCTGAGGT 2150
30 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCTGAG GTCAAGTTCA 2200
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35 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300
GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350
AAGCCCTCCC AGCCCCATC GAGAAAACCA TCTCCAAAGC CAAAGGGCAG 2400
45 CCCCAGAGAAC CACAGGTGTA CACCCTGCCC CCATCCCGGG AAGAGATGAC 2450
CAAGAACCAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCCAGCG 2500
50 ACATCGCCGT GGAGTGGGAG AGCAATGGGC AGCCGGAGAA CAACTACAAG 2550
ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600
GCTCACCGTG GACAAGAGCA GGTGGCAGCA GGGGAACGTC TTCTCATGCT 2650
60 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700
CTGTCTCCGG GTAAATGAGT GCGACGGCCC TAGAGTCGAC CTGCAGAAGC 2750
65 TTGGCCGCCA TGGCCCAACT TGTTTATTGC AGCTTATAAT GGTTACAAAT 2800

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5 CGATCGGGAA TTAATTCGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA 2950
AGAGGAACTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000
10 TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA 3050
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15 CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC 3150
AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAATCCGC 3200
20 CCAGTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT TTTTATTTAT 3250
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30 AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350
CCGCTTAATT AAGGCGCGCC ATTTAAATCC TGCAGGTAAC AGCTTGGCAC 3400
35 TGGCCGTCGT TTTACAACGT CGTGACTGGG AAAACCCTGG CGTTACCCAA 3450
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45 AATGGCGCCT GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTCA 3600
CACCGCATAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT 3650
50 AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG 3700
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TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC 3800
60 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA 3850
TGTTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTTGA 3900
65 CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTCCA AACTGGAACA 3950

ACACTCAACC CTATCTCGGG CTATTCTTTT GATTTATAAG GGATTTTGCC 4000
5 GATTTTCGGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAACG 4050
CGAATTTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100
10 ACAATCTGCT CTGATGCCGC ATAGTTAAGC CAACTCCGCT ATCGCTACGT 4150
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250
TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC 4300
20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTTT 4350
ATAGGTTAAT GTCATGATAA TAATGGTTTC TTAGACGTCA GGTGGCACTT 4400
TTCGGGGAAA TGTGCGCGGA ACCCCTATTT GTTTATTTTT CTAAATACAT 4450
30 TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA 4500
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35 TTCCCTTTTT TCGGCATTT TGCCTTCCTG TTTTGGCTCA CCCAGAAACG 4600
CTGGTGAAAG TAAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA 4650
CATCGAACTG GATCTCAACA GCGGTAAGAT CCTTGAGAGT TTTGCCCCG 4700
45 AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTTCTGCT ATGTGGCGCG 4750
GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA 4800
50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC 4850
TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG 4900
AGTGATAACA CTGCGGCCAA CTTACTTCTG ACAACGATCG GAGGACCGAA 4950
60 GGAGCTAACC GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050
65 ACCACGATGC CAGCAGCAAT GGCAACAACG TTGCGCAAAC TATTAAGTGG 5100

CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150

5 CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200

TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT 5250

10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300

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15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400

ATATATACTT TAGATTGATT TAAAACTTCA TTTTAAATTT AAAAGGATCT 5450

20 AGGTGAAGAT CCTTTTTGAT AATCTCATGA CCAAATCCC TTAACGTGAG 5500

25 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC 5550

TTGAGATCCT TTTTTTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC 5600

30 CACCGCTACC AGCGGTGGTT TGTTTGCCGG ATCAAGAGCT ACCAACTCTT 5650

TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCA ATACTGTCCT 5700

35 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5750

40 CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800

GATAAGTCGT GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA 5850

45 GGCGCAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG 5900

AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA 5950

50 AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG 6000

55 CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 6050

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA 6100

60 TTTTGTGAT GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA 6150

CGCGGCCTTT TTACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCACATGT 6200

65 TCCTTCCTGC GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC 6300
AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350
5 CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400
AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATTAA 6450
10 GGCACCCAG GCTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA 6500
15 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC 6550
GAATTAA 6557
20

(2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7305 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
35 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
40 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
50 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
60 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
65 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650
CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700
5 TTGGAACGCG GATTCCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA 750
GTCTATAGGC CCACCCCCTT GGCTTCGTTA GAACGCGGCT ACAATTAATA 800
CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA 850
15 CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC 900
ACCTCGGTTC TAAGCTTATC GATATGAAA AGCCTGAACT CACCGCGACG 950
20 TCTGTCGAGA AGTTTCTGAT CGAAAAGTTC GACAGCGTCT CCGACCTGAT 1000
GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050
GGCGTGGATA TGTCTGCGG GTAAATAGCT GCGCCGATGG TTTCTACAAA 1100
30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150
AGTGCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC 1200
35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCCC 1250
GCTGTTCTGC AGCCGGTCGC GGAGGCCATG GATGCGATCG CTGCGGCCGA 1300
TCTTAGCCAG ACGAGCGGGT TCGGCCATT CGGACCGCAA GGAATCGGTC 1350
45 AATACACTAC ATGGCGTGAT TTCATATGCG CGATTGCTGA TCCCCATGTG 1400
TATCACTGGC AAAGTGTGAT GGACGACACC GTCAGTGCGT CCGTCGCGCA 1450
50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGAAGTCCCC GAAGTCCGGC 1500
ACCTCGTGCA CGCGGATTTC GGCTCCAACA ATGTCCTGAC GGACAATGGC 1550
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60 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTGTATGG 1650
AGCAGCAGAC GACTTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG 1700
65 CGGCTCCGGG CGTATATGCT CCGCATTGGT CTTGACCAAC TCTATCAGAG 1750

CTTGGTTGAC GGCAATTTTCG ATGATGCAGC TTGGGCGCAG GGTCGATGCG 1800
5 ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATCGCC 1850
CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900
10 TAGTGAAAC CGACGCCCCA GCACTCGTCC GAGGGCAAAG GAATAGAGTA 1950
GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC 2000
15 CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA 2050
CAAATTTTAC AAATAAAGCA TTTTTTTCAC TGCATTCTAG TTGTGGTTTG 2100
20 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGGAATTAAT 2150
25 TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGGTTA 2200
GGTACCTTCT GAGGCGGAAA GAACCAGCTG TGAATGTGT GTCAGTTAGG 2250
30 GTGTGAAAG TCCCCAGGCT CCCAGCAGG CAGAAGTATG CAAAGCATGC 2300
ATCTCAATTA GTCAGCAACC AGGTGTGGAA AGTCCCCAGG CTCCCCAGCA 2350
35 GGCAGAAGTA TGCAAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC 2400
40 GCCCCTAACT CCGCCCATCC CGCCCCTAAC TCCGCCCAGT TCCGCCCAT 2450
CTCCGCCCCA TGGCTGACTA ATTTTTTTTA TTTATGCAGA GGCCGAGGCC 2500
45 GCCTCGGCCT CTGAGCTATT CCAGAAGTAG TGAGGAGGCT TTTTGGAGG 2550
CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGGAAC GGTGCATTGG 2600
50 AACGCGGATT CCCCCTGCCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT 2650
55 ATAGGCCAC CCCCTTGGCT TCGTTAGAAC GCGGCTACAA TTAATACATA 2700
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GTCCCCGAGC TCCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT 2950
5 GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG 3000
TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACGCGGCCTC 3050
10 GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGGA 3100
CGGATTTTAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTCGCAACT 3150
15 TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTTG GACAGGGTAC 3200
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30 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC 3450
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40 CCGCCATGGC CCAACTTGTT TATTGCAGCT TATAATGGTT ACAAATAAAG 3600
CAATAGCATC ACAAATTTCA CAAATAAAGC ATTTTTTTTCA CTGCATTCTA 3650
45 GTTGTGGTTT GTCCAAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT 3700
CGGGAATTAA TTCGGCGCAG CACCATGGCC TGAAATAACC TCTGAAAGAG 3750
50 GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCAGCT GTGGAATGTG 3800
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5 TTTTTTGGAG GCCTAGGCTT TTGCAAAAAG CTGTTAACAG CTTGGCACTG 4150

GCCGTCGTTT TACAACGTCG TGA CTGGGAA AACCTGGCG TTACCCAACT 4200

10 TAATCGCCTT GCAGCACATC CCCCCTTCGC CAGCTGGCGT AATAGCGAAG 4250

AGGCCCGCAC CGATCGCCCT TCCCAACAGT TGC GTAGCCT GAATGGCGAA 4300

15 TGGCGCCTGA TCGGTATTT TCTCCTTACG CATCTGTGCG GTATTTTACA 4350

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20 CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG 4450

25 CCCTAGCGCC CGCTCCTTTC GCTTTCTTCC CTTCTTTTCT CGCCACGTT 4500

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55 GACGGGCTTG TCTGCTCCCG GCATCCGCTT ACAGACAAGC TGTGACCGTC 5000

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60 GAGGCAGTAT TCTGAAGAC GAAAGGGCCT CGTGATACGC CTATTTTTTAT 5100

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65 CGGGGAAATG TCGCGGAAC CCCTATTTGT TTATTTTTTCT AAATACATT 5200

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5 ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTTCCGTGT CGCCCTTATT 5300

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10 GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGGTGACGA GTGGGTTACA 5400

TCGAACTGGA TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA 5450

15 GAACGTTTTTC CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT 5500

ATTATCCCGT GATGACGCCG GGCAAGAGCA ACTCGGTCGC CGCATACACT 5550

20 ATTCTCAGAA TGAATTGGT GAGTACTCAC CAGTCACAGA AAAGCATCTT 5600

25 ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 5650

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30 AGCTAACCGC TTTTTTGCAC AACATGGGGG ATCATGTAAC TCGCCTTGAT 5750

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35 CACGATGCCA GCAGCAATGG CAACAACGTT GCGCAAATA TTAAGTGGCG 5850

40 AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG 5900

GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5950

45 TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTG 6000

CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG 6050

50 ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT 6100

55 AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT 6150

ATATACTTTA GATTGATTTA AACTTCATT TTTAATTTAA AAGGATCTAG 6200

60 GTGAAGATCC TTTTGTATAA TCTCATGACC AAAATCCCTT AACGTGAGTT 6250

TTCTGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCAAA GGATCTTCTT 6300

65 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAAC AAAAAACCA 6350

CCGCTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACTCTTTT 6400
5 TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC 6450
TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACCGCCT 6500
10 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA 6550
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15 CGCAGCGGTC GGGCTGAACG GGGGGTTTCGT GCACACAGCC CAGCTTGGAG 6650
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20 CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA 6750
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30 TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAAAAC GCCAGCAACG 6900
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35 TTTCCTGCGT TATCCCCTGA TTCTGTGGAT AACCGTATTA CCGCCTTTGA 7000
GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050
TGAGCGAGGA AGCGGAAGAG CGCCCAATAC GCAAACCGCC TCTCCCCGCG 7100
45 CGTTGGCCGA TTCATTAATC CAGCTGGCAC GACAGGTTTC CCGACTGGAA 7150
AGCGGGCAGT GAGCGCAACG CAATTAATGT GAGTTACCTC ACTCATTAGG 7200
50 CACCCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT 7250
GTGAGCGGAT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300
55 ATTAA 7305
60

CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene
5 provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional
10 regulatory region.
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.
20
5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.
25
7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.
30
9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic
35 host.
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.
40
13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1
45 integrated into a chromosome of the host cell.

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the host cell of claim 11 so as to express the product gene and recovering the product from the host cell culture.
5
17. The method of claim 16 further comprising recovering the product from the culture medium.
- 10 18. The method of claim 16 wherein the selectable gene is an amplifiable gene and the splice donor site comprises an efficient splice donor sequence.
19. A method for producing a product of interest comprising culturing the host cell of claim 12 so as to express the product gene in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, and recovering the product.
15
20. A method for producing eukaryotic cells having multiple copies of a product gene comprising transforming eukaryotic cells with the DNA construct of claim 5, growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene.
20
21. The method of claim 20 further comprising recovering from the selected cells the product of interest.
25

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FIG. 1A

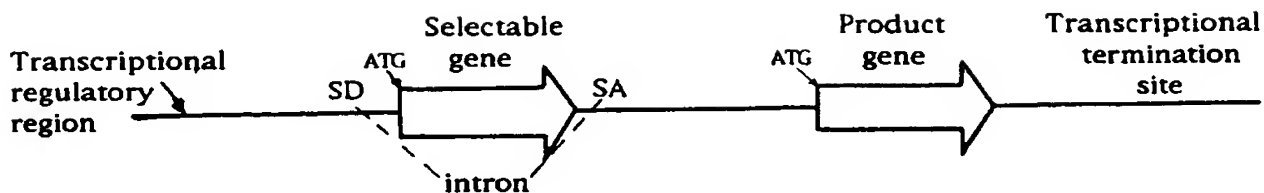


FIG. 1B

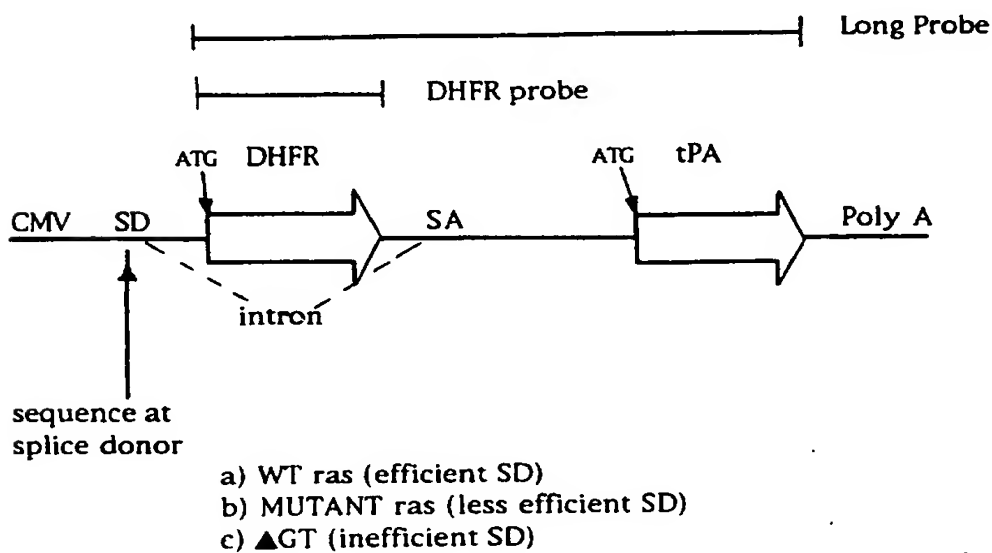
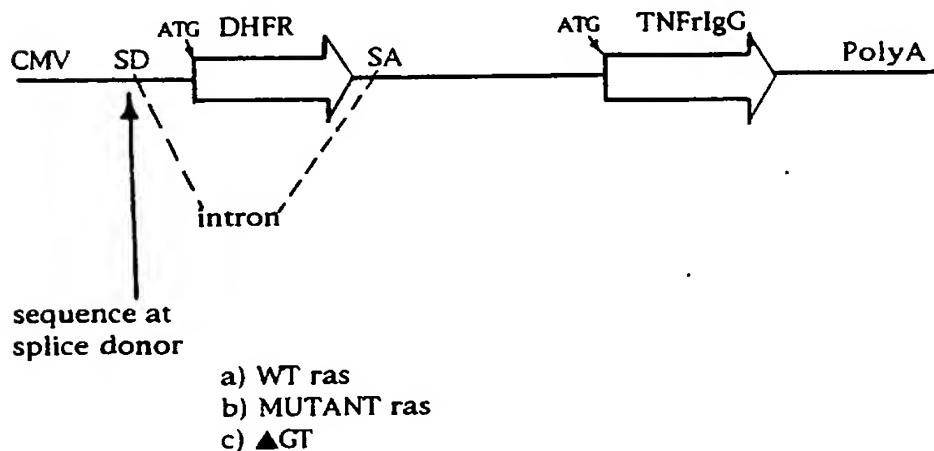


FIG. 1C



SUBSTITUTE SHEET (RULE 26)

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FIG. 1D

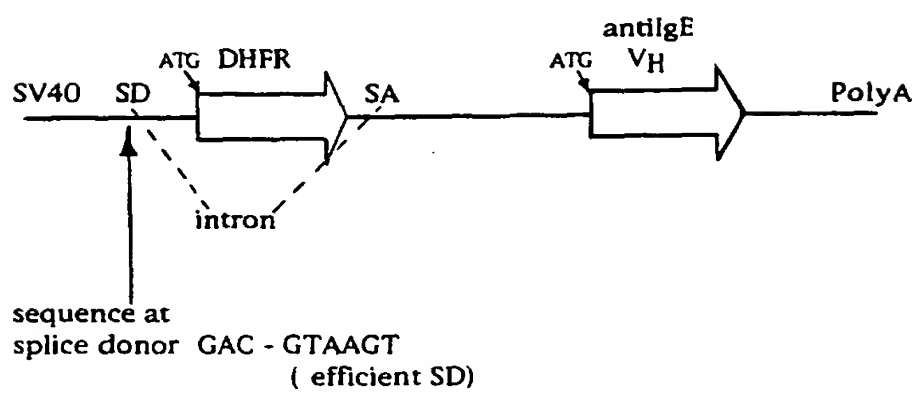
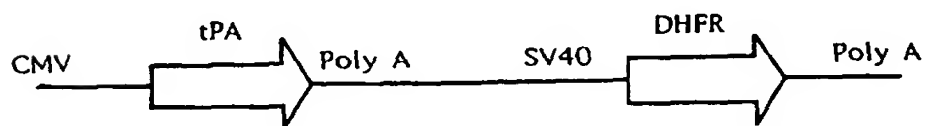


FIG. 2



SUBSTITUTE SHEET (RULE 26)

FIG. 3A

[illegible]

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FIG. 3B

```

401  GGTGTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA TTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA
    CCAAAACCGT CATGTAGTTA CCGGCACCTA TCGCCAAACT GAGTGCCCT AAAGTTTCA AGGTGGGTA ACTGCAGTTA CCTCAACA AAACCGTGTG

    rsaI      pleI      maeII      hinII/acyI      nlaIV
    csp6I      aciI      hinfI      ahaII/bsaHI      hgiCI
    bsmAI      aatII      bsmAI      aatII      banI

501  AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCGCC CCATTGACGC AAATGGCGG TAGGCTGTA CGGTGGAGG TCATATATAG CAGAGCTCGT
    TTTAGTTGCC CTGAAGGTT TTACAGCATT GTTGAGCGG GGTAAGTGG TTTACCCGCC ATCCGCACAT GCCACCTCC AGATATATTC GTCTCGAGCA

    rsaI      hgaI      mnlI      csp6I      mnlI
    aciI      aciI      mnlI      csp6I      mnlI

    haeIII/palI
    mcrI
    eagI/xmaIII/ecI XI
    eaeI
    cfrI
    fnu4HI
    aciI
    thaI
    fnuDII/mvni

    sau96I      sacII/sstII      nspBII
    avaiI      kspI      scrFI
    asuI      dsaI      nciI
    nlaIV      scrFI      bglI      bslI      mspI
    aciI      sau3AI      mnlI      bstUI
    nciI      mboI/ndeII(dam-) hpaII
    mspI      mboI/ndeII(dam-) hpaII
    hpaII      dpnI(dam+) bsaJI dsav
    dsav      dpnII(dam-) bsh1236I
    cauII      alwI(dam-) aciI cauII

601  TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCAGCT GTTTGACCT CCATAGAAGA CACCGGACC GATCCAGCCT CCGCGCCCGG GAACGGTGCA
    AATCACTTGG CAGTCTAGCG GACCTCTGCG GTAGTGCGA CAAACTGGA GGATCTTCT GTGCGCCCTGG CTAGGTGCGA GCGCGCCGCC CTTGCCACGT

    esp3I
    scrFI
    mvaI bsmAI
    ecorII
    dsav
    bstNI hinII/acyI
    apyI(dcm+)
    sau3AI gsui/bpmI
    mboI/ndeII(dam-)
    dpnI(dam+) hgaI foki
    dpnII(dam-) ahaII/bsaHI
    mnlI

```

FIG. 3C

tfII
 aciI
 thal hinfI
 fnuDII/mvnI
 bstUI
 bsh1236I
 701 TTGGAACGGG GATTCCCGG GCCAAGAGTG CTGTAAGTAC CGCTATAGA GCGATAAGAG GATTTTATCC CCGTGCCAT CATGGTTGGA CCATTGAACT
 AACCTTGGC CTAAGGGCA CGGTTCTCAC GACATTTCAT GCGATATCT CCGATATCTC CTAAATAGG GCGCAGGTA GTACCAAGCT GGTAACCTGA
 fnu4HI
 bbvI
 nspBII
 aciI
 nlaIII
 taqI
 thal
 fnuDII/mvnI
 bstUI
 bsh1236I
 mluI
 bsrBI
 aflIII
 rsaI
 aciI
 xmnI
 csp6I
 mnlI
 ddeI
 asp700
 scaI
 801 GCATCGTCCG CGTGTCCTCA AATATGGGA TTGGCAAGAA CGGACACCTA CCCTGCCCTC CGCTCAGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC
 CGTAGCAGG GCACAGGGT TTATACCCCT AACCGTTCTT GCCTCTGGAT GGGACGGAG GCGAGTCTT CCGCAAGTTC ATGAAGGTTT CTTACTGGTG
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI(dcm+)
 sexAI
 tfII
 hinfI
 alwNI
 hphI
 901 AACCTCTTCA GTGGAAGTA AACAGAATCT GGTGATTATG GGTAGGAAA CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAA
 TTGGAGAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAAATTTCTT GTCTTAATTA
 aluI
 sstI
 sacI
 hgiJII
 hgiAI/aspHI
 eclI36II
 bsp1286
 bsiHKA
 bmyI
 banII
 bslI
 mnlI
 ddeI
 1001 ATAGTTCTCA GTAGAGACT CAAAGAACA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGAG CTTAAGACT TATTGAACAA CCGGAATTGG
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCCTC GAGTAAAGA ACGGTTTCA AACCTACTAC GGAATTTCTG ATAACTTCTT GGCCTTAACC
 mspl
 hpaII
 bsaI

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FIG. 3D

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      haeIII/palI
      haeI
      nlaIII
      scrFI      mvaI      sau3AI
      mvaI      ecorII      mboI/ndeII(dam-)
      dsav      tfil      dsav      pleI      dpnI(dam+)
      bstNI      nlaIII      bstNI      ddeI      dpnII(dam-)
      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      TGGTATGTC      TGGTATGTC      TGGTATGTC      TGGTATGTC      TGGTATGTC      TGGTATGTC
      accI      nlaIII      mnlI      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      1101 CAAGTAAAGT AGACATGGTT TGGTATGTC GAGCAGTTC TGGTATGTC TGGTATGTC TGGTATGTC TGGTATGTC TGGTATGTC
      GTTCATTCA TCTGTACCA ACCTATCAGC CTCCGTCAAG ACAATGGTC CTCCGTCTT TCTGTATGTC TCTGTATGTC TCTGTATGTC TCTGTATGTC
      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI
      scrFI      mvaI      ecorII      dsav      bstNI      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      1201 GCAGGAATT GAAAGTGACA CGTTTTCCT AGAATGAT TTGGGAAAT ATAACTCTT CCAGATATC CCAGATATC CCAGATATC CCAGATATC
      CGTCCCTAAA CTTTCACTGT GCAAAAAGGG TCTTTAACTA AACCCCTTA TATTGGAGA GGTCTTATG GGTCTTATG GGTCTTATG GGTCTTATG
      apoI      maeIII      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI
      scrFI      mvaI      ecorII      dsav      bstNI      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      1301 AAAGGCATCA AGTATAGTT TGAAGTCTAC GAGAGAAAG ACTAACAGGA AGATGCTTC AAGTCTCTG CTCCCTCTC AAAGCTATGC ATTTTATAA
      TTCCCGTAGT TCATATTCAA ACTTCAGATG CTCTTCTTTC TGATGTCCT TCTACGAAAG TTCAAGAGAC GAGGGAGGA TTTCGATACG TAAAAATATT
      sfanI      mboII      accI      mboII      mboII      mboII      mboII      mboII      mboII
      scrFI      mvaI      ecorII      dsav      bstNI      apyI(dcm+)      hinfi      apyI(dcm+)      hinfi      maeIII      alwI(dam-)
      1401 GACCATGGGA CTTTTCCTGG CTTTAGACCC CTTTGGCTTC GTTAGAAGC GGTACAAAT ATACATAAC CTTATGTATC ATACATAG ATTTAGGTGA
      CTGGTACCCCT GAAACGACC GAATCTGGG GGAACCGAAG CAATCTGGG CCGATGTAA TATGTATTG GAATACATAG TATGTATC TAAATCCACT
      fnu4HI      aciI      thai      fnuDII/mvnI      tru9I      bstUI      mseI      bsh1236I      aseI/asnI/vsPI
      nlaIII      styI      ncoI      dsal      bsalI      styI      bsalI      styI      bsalI
      maeIII      hphI

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[illegible]

FIG. 3F

bspMI sau96I
 nlaIV haeIII/palI
 hgiCI asuI rsal
 banI eco109I/draII
 bsp1286 alwNI csp6I ddeI
 bmyI 1901 GGGGGCACCT GCCAGCAGGC CCGTGTACTTC TCAGATTTTCG TGTGCCAGTG CCCCGAAGGA TTGCTGGGA AGTCTGTGA AATAGATACC AGGGCCACGT
 CCCCCGTGGA CGGTGCTCCG GGACATGAAG AGTCTAAAGC ACACGGTCAC GGGCTTCCT AACGACCCT TCACGACACT TTATCTATGG TCCCGGTGCA

pmlI pmlI
 eco72I eco72I
 sau96I sau96I
 asuI maeII
 scrFI scrFI
 mvaI haeIII/palI
 ecoRII
 dsav
 bstNI bsaAI
 bsaJI bbrPI
 apyI[dcm+]

bsp1286
 bmyI bsrI bcgI
 2001 GCTACGAGGA CCAGGGCATC AGCTACAGGG GCACGTGGAG CACAGCGGAG AGTGGCGCG AGTGACACCAA CTGGAACAGC AGCGCGTTGG CCCAGAAGCC
 CGATGCTCCT GGTCCCGTAG TCGATGTCCC CGTGCACCTC GTGTGCGCTC TCACCGCGG TCACGTGGTT GACCTTGTGG TCGCGCAACC GGTCTTTCGG

hinPI hinPI
 hhaI/cfoI
 nlaIV hgiAI/aspHI
 narI bsp1286
 kasi bsiHKAI
 hinII/acyI
 pmlI hgiAI/aspHI
 eco72I
 bsaAI bsp1286
 bbrPI bsiHKAI
 asuI sfaNI scfI bsp1286 bmyI acil banI alw44I/snoI
 mnli apyI[dcm+] alul bmyI maeII nspBII ahaII/bsaHI bsrI fnu4HI asuI bsli
 2001 GCTACGAGGA CCAGGGCATC AGCTACAGGG GCACGTGGAG CACAGCGGAG AGTGGCGCG AGTGACACCAA CTGGAACAGC AGCGCGTTGG CCCAGAAGCC
 CGATGCTCCT GGTCCCGTAG TCGATGTCCC CGTGCACCTC GTGTGCGCTC TCACCGCGG TCACGTGGTT GACCTTGTGG TCGCGCAACC GGTCTTTCGG

scrFI scrFI
 mvaI mvaI
 ecoRII ecoRII
 dsav dsav
 bstNI bsaAI
 bsaJI apyI[dcm+]

hinII/acyI
 haeIII/palI
 nspBII haeI hgaI bstXI
 scfI aciI mnli ahaII/bsaHI
 2101 CTACAGCGGG CGGAGGCCAG AGCCCATCAG GCTGGGCCCTG GGAACACCA ACTACTGCAG AAACCCAGAT CGAGACTCAA AGCCCTGGTG CTACGTCTTT
 GATGTCGCCC GCCTCCGGTC TCGGGTAGTC CGACCCGGAC CCCTGGTGT TGATGACGTC TTGGGTCTA GCTCTGAGTT TCGGACCAC GATGCAGAAA

pleI pleI scrFI
 bsmAI mvaI
 taqI[dam-] ecoRII
 sau3AI hinfI dsav
 mboI/ndeII[dam-] bstNI
 apyI[dcm+] tru9I
 bsaJI maeII mseI

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FIG. 3G

[illegible]

FIG. 3H

mspl
 hpall
 bsli
 scrFI
 ncil
 dsav
 nlaIV
 acil
 bsrBI
 cauII
 mnlI
 bsli
 2601 CCATCTTTGC CAAGCACAGG AGGTGCCCCG GAGAGCGGTT CCTGTGCGGG GGCATACTCA TCAGCTCCTG CTGGATTCTC TCTGCCGCC ACTGCTTCCA
 GGTAGAAACG GTTCGTGTCC TCCAGCGGGC CTCTGCCCAA GGACACGCC CCCTATGAGT AGTCGAGGAC GACCTAAGAG AGACGGCGGG TGACGAAGGT
 scrFI
 mval
 ecorII
 dsav
 bstNI
 apyl{dcm+}
 aluI
 alwNI
 tfil
 hinfI
 fnu4HI
 acil
 2701 GGAGAGGTTT CGCCCCACC ACCTGACGGT GATCTTGGC AGAACATACC GGGTGTGCC TGGCGAGGAG GAGCAGAAAT TTGAAGTCCA AAATACATT
 CCTCTCCAAA GCGCGGGTGG TGGACTGCCA CTAGAACCAG TCTTGTATGG CCCACCAGGG ACCGCTCCTC CTCGTCTTTA AACTTCAGCT TTTTATGTAA
 mnlI
 acil
 bsli
 hphI{dam-}
 cauII
 asuI
 mnlI
 apoI
 taqI
 scrFI
 mval
 ecorII
 dsav
 bstNI
 apyl{dcm+}
 bsli
 ncil
 mspl
 hpall
 dpnI{dam+}
 dpnII{dam-}
 dsav
 avall
 cauII
 asuI
 mnlI
 apoI
 taqI
 scrFI
 pstI
 fnu4HI
 bbvI
 pvuII
 fnu4HI
 bbvI
 hinPI
 bsgI
 aluI
 hhai/cfoI
 nspBII
 2801 GTCCATAAGG AATTCGATGA TGACACTTAC GACAATGACA TTGCGTGTCT GCAGCTGAAA TCGGATTCTG CCCGCTGTGC CCAGGAGAGC AGCGTGTGTC
 CAGGTATTCC TTAAGCTACT ACTGTGAATG CTGTTACTGT AACGCGACGA CGTCGACTTT AGCCTAAGCA GGGCGACACG GGTCTCTCTG TCGCACACAG
 ecorI
 apoI
 taqI
 scrFI
 mval
 ecorII
 dsav
 bstNI
 bspI286
 nspBII
 bsaJI
 bbvI
 fnu4HI
 sau96I
 acil
 2901 GTCCATAAGG AATTCGATGA TGACACTTAC GACAATGACA TTGCGTGTCT GCAGCTGAAA TCGGATTCTG CCCGCTGTGC CCAGGAGAGC AGCGTGTGTC
 CAGGTATTCC TTAAGCTACT ACTGTGAATG CTGTTACTGT AACGCGACGA CGTCGACTTT AGCCTAAGCA GGGCGACACG GGTCTCTCTG TCGCACACAG

FIG. 31

[illegible]

SUBSTITUTE SHEET (RULE 26)

SUBSTITUTE SHEET (RULE 26)

BNSDOCID: <WO__9604391A1_|_>

BNSDOCID: <WO__9604391A1_1_>

FIG. 3N

FIG. 3N

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4801  GCCGATTTCG  GCCTATTGGT  TAAAAAATGA  GCTGATTTTAA  CAJAAATTTA  ACGCGAATTT  TAACGATTTA  TTAACGTTTA  CAATTTTATG  GTGCACCTCTC
      haeIII/palI      aluI      mseI      apoI      apoI      bsh1236I      sspl mseI
      tru9I      tru9I      mseI      bstUI      mseI      tru9I      psp1406I
      mseI      tru9I      mseI      apoI      apoI      bsh1236I      sspl mseI
      tru9I      apoI      tru9I      apoI      apoI      maeII      psp1406I
      maeII/mvnI      maeII      maeII      apoI      apoI      maeII      psp1406I
      bsiHKAI      bmyI      ddeI      apoI/snoI      alw44I/snoI
      hgiAI/aspHI      bsp1286      bsiHKAI      bmyI      ddeI      apoI/snoI      alw44I/snoI      GTGCACCTCTC
      CCGCTAAAGC  CGGATAACCA  ATTTTCTACT  CGACTAAATT  GTTTTAAAT  TCGCGTTAAA  ATTGTTTAT  AATTGCAAT  GTTAAATAC  CACGTGAGAG
      hinPI      hhaI/cfoI      thaI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      rsaI      fnu4HI      tru9I      mseI      aciI      aciI      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      csp6I      sfaNI      mseI      mseI      mseI      mseI      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
4901  AGTACAATCT  GCTCTGATGC  CGCATAGTTA  AGCCAATCC  GCTATCGCTA  CGTGACTGGG  TCATGGCTGC  GCCCGGACAC  CCGCCAAAC  CCGCTGACGC
      TCATGTTAGA  CGAGACTACG  GCGTATCAAT  TCGGTTGAGG  CGATAGCGAT  GCACTGACCC  AGTACCGACG  CCGGGCTGTG  GCGGCTTGTG  GCGGACTGCG
      scrFI      nciI      dsav      foki      cauII      aciI      cauII      aluI      nlaIII      mnlI      hphI
      dsav      foki      cauII      aciI      cauII      aluI      nlaIII      mnlI      hphI
5001  GCGCTGACGG  GCTTGCTGTC  TCCCGGCATC  CGCTTACAGA  CAAGCTGTGA  CCGTCTCCGG  GAGCTGCATG  TGTCAGAGGT  TTTCACCGTC  ATCACCGAAA
      drdI      drdI      cauII      aciI      cauII      aluI      nlaIII      mnlI      hphI      hphI
      GCGGACTGCC  CGAACAGACG  AGGCGCGTAG  GCGAATGTCT  GTTCGACACT  GGCAGAGGCC  CTCGACGTAC  ACAGTCTCCA  AAAGTGGCAG  TAGTGGCTTT
      thaI      fnuDII/mvnI      bstUI      bsh1236I      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
5101  CGCGCGAGGC  AGTATTCTTG  AAGACGAAAG  GGCCTCGTGA  TACGCCATAT  TTTATAGGTT  AATGTCATGA  TAATAATGGT  TTCTTAGACG  TCAGGTGGCA
      bsh1236I      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      bsh1236I      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      GCGCGCTCCG  TCATAAGAAC  TTCTGCTTTC  CCGGAGCACT  ATGCGGATAA  AAATATCCAA  TTACAGTACT  ATTATTACCA  AAGAATCTGC  AGTCCACCGT
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      bsh1236I      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI      aciI hgaI
      hinPI      hhaI/cfoI      thaI mnlI      fnuDII/mvnI      bstUI      nspBII bsh1236I
      fnuDII/mvnI      fnu4HI      maeIII      maeII bsrI      nlaIII hhaI/cfoI      bstUI      nspBII bsh1236I
      fnu4HI      maeII      bsaAI tth111I/aspI bbvI      aciI
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FIG. 30

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nlaIV
aciI
thai
fnuDII/mvni
bstUI
bsh1236I
hinPI
hhaI/cfoI

5201 CTTTTCGGG AAATGTGCGC GGAACCCCTA TTTGTTTATT TTTCTAAATA CATTCAAATA TGTATCCGCT CATGAGACAA TAACCTGTAT AAATGCTTCA
GAAAGCCCC TTTACAGCGC CTTGGGGAT AAACAAATAA AAAGATTTAT GTAAAGTTTAT ACATAGCGCA GTACTCTGTT ATTGGGACTA TTTACGAAGT

        rcaI
        bspHI
        bsrBI bsmAI
        aciI nlaIII
        fnu4HI
        aciI
        hphI

5301 ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTGCCC TTATTCCCTT TTTTGGCGCA TTTTGCTTTC CTGTTTTTGC TCACCCAGAA
TATTATAACT TTTTCTTCT CATACTCATA AGTTGTAAAG GCACAGCGG AATAAGGGA AAAACGCCGT AAAACGGAAG GACAAAAACG AGTGGGTCTT

        hgiAI/aspHI
        bsp1286
        sau3AI bsiHKAI
        mboI/ndeII[dam-]
        dpnI[dam+] bmyI
        dpnII[dam-]
        eco57I apaII/snoI
        hphI sfaNI mboII[dam-] alw44I/snoI maeIII taqI alwI[dam-] aciI bstyI/xhoII

5401 ACCTGGTGA AAGTAAAGA TGCTGAAGAT CAGTTGGTG CACGAGTGG TTACATCGAA CTGGATCTCA ACACGGTAA GATCCTTGAG AGTTTTGCGC
TGCGACCACT TTCACTTCT ACGACTTCTA GTCAACCCAC GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTCGCCATT CTAGGAATC TCAAAAGCGG

        scrFI
        nciI
        mspI
        hpaII
        dsav
        hinII/acyI
        hgaI cauII
        aciI
        bsgI mcrI fnu4HI

5501 CCGAAGAAGC TTTTCCAATG ATGAGCACTT TTAAGTTCT GCTATGTGC GCGGTATTAT CCGTGATGA CGCGGGCAA GAGCAACTCG GTCGCCGCAT
GGCTTCTTGC AAAAGGTAC TACTCGTGAA AATTTCAGA CGATACACCG CGCCATAATA GGGCACTACT GCGGCCCGTT CTCGTTGAGC CAGCGGCGTA

        rsaI
        csp6I bsrI
        scaI hphI maeIII sfaNI foki nlaIII
        ddeI
        fnu4HI
        bbvI nlaIII

5601 ACACCTATTCT CAGAAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC TGCCATAACC
TGTCATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTCCAG TGCTTTTTCG TAGAATGCCT ACCGTACTGT CATCTCTTA ATACGTCACG ACGGTATTGG

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FIG. 3P

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sau96I
avaII
sau3AI asuI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
pvuI/bspCI
mcrI mnlI
aluI aciI
nlaIII maeIII
sau3AI maeIII
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
nlaIII alwI(dam-)
5701 ATGAGTGATA ACACGTGCGC CAACCTACTT CTGACAACGA TCGGAGGACC GAAGGAGCTA ACCGTTTTT TGCACAACAT GGGGATCAT GTAACTCGCC
TACTCACTAT TGTGAGCGCG GTTGAATGAA GACTGTGCT AGCCTCCTGG CTTCCTCGAT TGGCGAATAA ACGTCTTGA CCCCTAGTA CATTGAGCGG
haeIII/palI
eaeI
cfrI
fnu4HI
aciI
mspI
sau3AI nlaIV
mboI/ndeII(dam-) aluI
dpmI(dam+) hpaII
dpmII(dam-) bsaWI
5801 TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACAAA CGACGAGCGT GACACACAGA TGCCAGCAGC AATGGCAACA ACCTTGCGCA AACTATTAC
AACTAGCAAC CCTTGCCCTC GACTTACTTC GGTATGTTT GCTGCTCGCA CTGTGTGCT ACCTGCTGCT TACCGTTGT TGCAACGCGT TTGATAATTG
hinPI
hhaI/cfoI
mstI
aviII/fspI bsrI
maeII
psp1406I
fnu4HI
sfaNI
bbvI
sau96I
bglI
sau96I
haeIII/palI
hinPI asuI mspI
hhaI/cfoI hpaII
5901 TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG AGCGGATAA AGTTGCAGGA CCACCTCTGC GCTCGCCCT TCCGGCTGGC
ACCGCTTGAT GAATGAGATC GAAGGCGCGT TGTTAATTAT CTGACCTACC TCCGCTATT TCAACGTCCT GGTGAAGACG CGAGCCGGA AGGCCGACCG
mspI
hpaII
cfrI
nlaIV hphI
gsuI/bpmI
thaI
fnuDII/mvni
bstuI
sau96I
bsmaI aciI
fnu4HI nlaIV
bsaI bsh1236I bbvI bsrI asuI
mnlI
6001 TGGTTTATTG CTGATAAATC TGGAGCGGT GAGCGTGGT CTGCGGTAT CATTGCAGA CTGGGCGCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT
ACCAATAAC GACTATTAG ACCTCGCCA CTGCGACCA GAGCGCCATA GTACGTCGT GACCCCGTC TACCATTCCG GAGGCGATAG CATCAATAGA
ddeI
sau3AI nlaIV
mboI/ndeII(dam-)
dpmI(dam+) hgiCI
dpmII(dam-) bsrI mnlI maeIII
tru9I
mstI
6101 ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAATAG ACAGATCGT GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACGTGTCG ACCAAGTTTA
TGTGCTGCC CTGAGTCCGT TGATACCTAC TTGCTTATC TGTCTAGCA CTCTATCCAC GGAGTACTA ATTGTAACC ATTGACAGTC TGGTTCAAT

```


FIG. 3Q

6201 CTCATATATA CTTTAGATTG ATTTAAAACT TCATTTTAA TTTTAAAGGA TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAAT CCCTTAACGT
GAGTATATAT GAAATCTAAC TAAATTTTGA AGTAAATAAT AAAATTTTCT AGATCCACTT CTAGGAAAAA CTATTAGAGT ACTGGTTTTA GGGAAATTGCA

6301 GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAGGATC TTCTTGAGAT CCTTTTTC TCGCGGTAAT CTGCTGCTTG CAAACAAAAA
CTCAAAAGCA AGGTGACTCG CAGTCTGGGG CATCTTTTCT AGTTTCTTAG AAGAACTCTA GGAATAAAAG ACGCGCATTA GACGACGAAC GTTTGTTTTT

6401 AACCAACCGT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTCCGA AGTAACTGG CTTCAGAGAG CGCGAGATAC CAAATACTGT
TTGGTGGCGA TGGTCGCCAC CAAACAAACG GCGTAGTTCT CGATGTTTGA GAAAAAGGCT TCCATTGACC GAAGTCGTCT CGCGTCTATG GTTTATGACA

6501 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC CGCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT
GGAGATCAC ATCGGTCATCA ATCCGGTGGT GAAGTTCTTG AGACATCGTG GCGGATGTAT GGAGCGAGAC GATTAGGACA ATGSTACCG ACGACGGTCA

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FIG. 3R

```

6601 GCGGATAAGT CGTGCTTAC CCGGTTGGAC TCAAGACGAT AGTTACCGGA TAAGCGCAG CCGTCGGCT GAACGGGGGG TTCTGTGCACA CAGCCAGCT
CCGCTATTCA GCACAGAATG GCCCAACCTG AGTCTGCTA TCAATGGCCT ATTCCGGCTC GCCAGCCGA CTTCGCCCC CTTGCCCTTC AAGCACGTGT GTCGGGTGCGA

6701 TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA GAAAGCGCCA CGCTTCCGA AGGAGAAAG GCGACAGGT ATCCGGTAAG
ACCTCGCTTG CTGGATGCG CTTGACTCTA TGGATGTGC ACTCGTAACT CTTTCGGGT CGGAAGGCT TCCCTCTTC CGCCTGTCCA TAGGCCATTC

6801 CGGCAGGTC GGAACAGGAG AGCGACGAG GGAGCTTCCA GGGGAAACG CCTGGTATCT TTATAGTCT GTCCGGTTTC GCCACCTCTG ACTTGAGGCT
GCGGTCCCAG CCTGTCTCTC TCCTGTCTC CTCTGAAGGT CCCCTTTGC GGACCATAGA AATATCAGGA CAGCCCAAAG CGGTGGAGAC TGAACCTCGA

6901 CGATTTTGT GATGCTCGTC AGGGGGCGG AGCCTATGGA AAAACGCCAG CAACCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCT TTTGCTCACA
GCTAAAAACA CTACGAGCAG TCCCCCGCC TCGGATACCT TTTTGGGCTC GTTGGCGCGG AAAAATGCCA AGGACCGGA AACGAGTGT

7001 TGTTCTTTCC TCGGTTATCC CCTGATTCTG TGGATAACCG TATTACGCC CTTTACCGCT TTTGAGTGAG CTGATACCG TCGCCGCGAG CGAAGCAGCG AGCGAGCGA
ACAAGAAAGG ACCTAATAGG GGAATAAGAC ACCTATTGGC ATAATGGCGG AACTCACTC GACTATGGG AGCGGCGTCG GCTTGTGGC TCGGCTCGCT

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FIG. 3S

```

          thal
          fnuDII/mvnI
          bstUI
          bsh1236I
          hinPI
          hhAI/cfoI
          thal
          fnuDII/mvnI
          bstUI haeIII/palI aluI
          bsh1236I tru9I pvuII
          bslI eaeI tfil aseI/asnI/vspI
          aciI cfrI hinfi msel nspBII bsrI
          CCGCGCGTGG GCCGATTTCAT TAATCCAGCT GGCACGACAG GTTCCCGAC
          CAGTCACTCG CTCCTTCGCC TTCTCGCGGG TTAGGCTGTA ATTAGGTGCA CCGTGTGTC CAAAGGGCTG
          7101

          sapi hinPI
          mboII hhaI/cfoI
          earI/ksp632I
          mnII aciI haeII
          CAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC
          CCGCGCGTGG GCCGATTTCAT TAATCCAGCT GGCACGACAG GTTCCCGAC
          CAGTCACTCG CTCCTTCGCC TTCTCGCGGG TTAGGCTGTA ATTAGGTGCA CCGTGTGTC CAAAGGGCTG
          7201

          hinPI tru9I
          hhaI/cfoI msel maeIII
          CAGGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG
          ACCTTTTCGCC CGTCACTCGC GTTGGGTTAA TTACACTCAA TGGAGTGAGT AATCCGTGGG GTCCGAAATG TGAATAACGA AGGCCGAGCA TACAACACAC
          7301

          aciI
          bsrBI aluI nlaIII
          GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT TACGAATTAA
          CTTAACACTC GCCTATTGTT AAAGTGTGTC CTTTGTGCGAT ACTGGTACTA ATGCTTTAATT
          7401

```

>length: 7360

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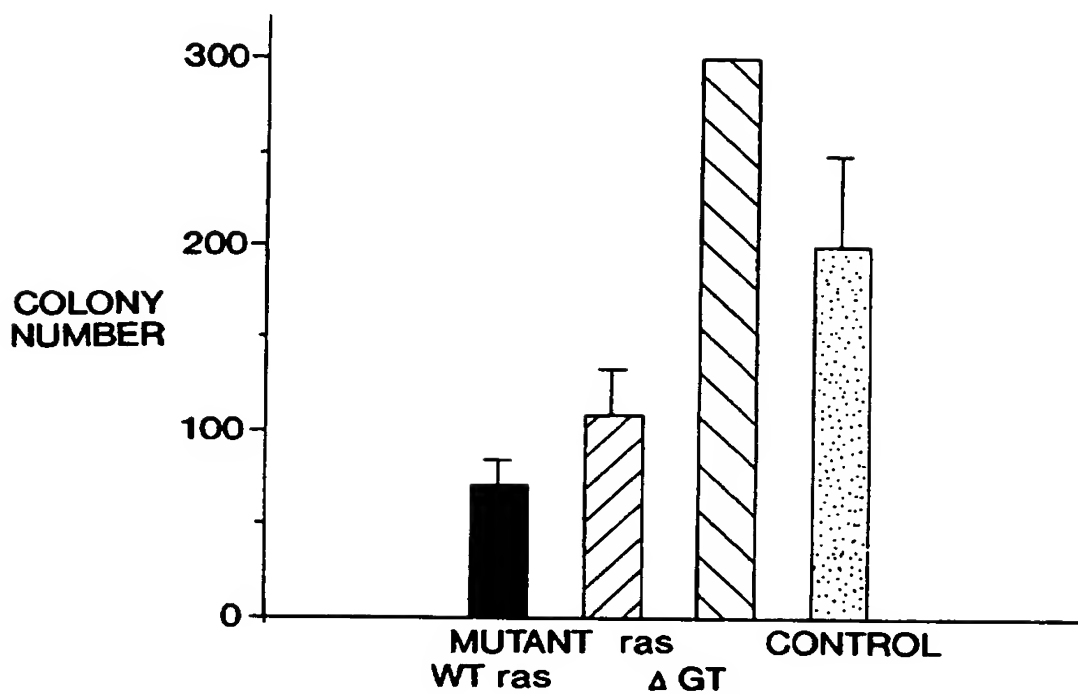


FIG. 4

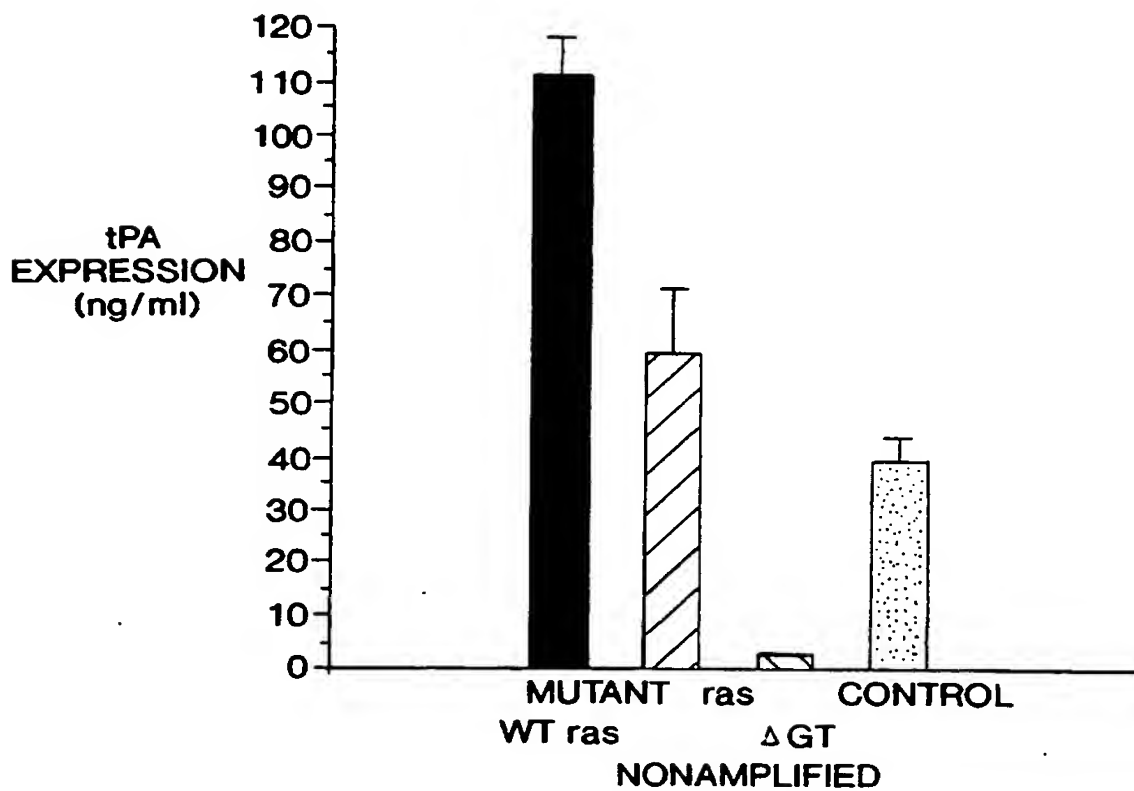


FIG. 5A

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FIG. 10A

```

1  TTCCAGCTCG CCCGACATG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
   AAGCTGAGC GGGCTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGGTATATA CCTCAAGGCG CAATGTATTG
   tagl          rmaI      tru9I          bslI          aciI maeIII
                maeI      mseI          speI      aseI/asnI/vspI
                speI      aseI/asnI/vspI
201 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
   AACTGCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGCT TCATGCGGGG GATNACTGCA GTTACTGCCA
   maeII          hinII/acyI          ahaII/bsaHI          aatII          maeII          maeIII
                hinII/acyI          ahaII/bsaHI          aatII          maeII          maeIII
201 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
   AACTGCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGCT TCATGCGGGG GATNACTGCA GTTACTGCCA
   maeII          hinII/acyI          ahaII/bsaHI          aatII          maeII          maeIII
                hinII/acyI          ahaII/bsaHI          aatII          maeII          maeIII
301 AAATGGCCCG CCTGGCATTA TGCCGAGTAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC
   TTTACCGGGC GGACCGTAAT ACGGTCATG TACTTGAATA CCTGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATAATGG TACCACCTAGC
   aciI          bglI dsav          sau96I          bstNI          bsrI nlaIII          styI          ncoI          dsaI hphI aciI
                bglI dsav          sau96I          bstNI          bsrI nlaIII          styI          ncoI          dsaI hphI aciI

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FIG. 9R

```

        thal
        fnuDII/mvnI
        bstUI
        bsh1236I
        hinPI
        hhai/cfoI
        thal
        fnuDII/mvnI
        bstUI
        bsh1236I haeIII/palI      tru9I   aluI
        bsII    eaeI  tfII aseI/asnI/vspI   pvuII
        aciI    cfrI  hinI mseI  nspBII
        6301 AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT AGCAAAACCG CCTCTCCCCG CGGCTTGGCC GATTCAATTAA TCCAGCTGGC ACCACAGGTT TCCCGACTGG
        TCACTCGCTC CTTCGCCCTC TCGCGGCTTA TCGGTTTGGC GGAGAGGGGC GCGCAACCGG CTAAGTAATT AGGTGACCG TGCTGTCCAA AGGGCTGACC
        bsrI

        scrFI
        mvaI
        ecorII
        dsav
        nlaIV bstNI
        hgiCI apyI[dcn+]      mspI
        banI bsaJI           hpaII
        6401 AAAGCGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCAATTA GGCACCCCGAG GCTTTACACT TTATGCTTCC GGTGCGTATG TTGTGTGGAA
        TTTGCCCCGT CACTCGCGTT GCGTTAATTA CACTCAATGG AGTGAGTAAT CCGTGGGGTC CGAAATGTGA AATACGAAGG CGAGCATAC AACACACCTT

        tru9I
        mseI
        aseI/asnI/vspI
        xnnI
        asp700
        6501 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC GAATTAA
        AACACTCGCC TATTGTTAAA GTGTGCTCTT TGTGATACT GGTACTAATG CTTAATT

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>length: 6557

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FIG. 9Q

```

                    hinPI          hhaI/cfoI          hhaII          aciI          bsaWI          fnu4HI
                    mspI          hpaII          bslI          bsaWI          aciI          fnu4HI
5901 AGCGAAGCAGC CTACACCGAA CTGAGATACC TACAGGTGA GCATTGAGAA AGCGCCACGC TTCCCGAAGG GAGAAGCGG GACAGGTATC CGGTAAGCGG
TCGCTTGCTG GATGTGGCTT GACTCTATGG ATGTCGCACT CGTAACTCTT TCGCGGTGGG AAGGCTTCC CTCTTTCCGC CTGTCCATAG GCCATTCCGC

                    scrFI          mvaI          ecorII          mvaI          dsaV          ecorII          bstNI          dsaV
                    hinPI          mnlI          hhaI/cfoI          aluI          apyI[dcM+]          apyI[dcM+]          bstNI          dsaV
6001 CAGGTCGGA ACAGGAGAGC GCACGAGGA GCTTCCAGGG GGAACGCCT GGTATCTTTA TAGTCTGTTC GGGTTTCGCC ACCTTGACT TGAGCGTGA
GTCCAGCCT TGTCTCTCG CGTCTCCCT CGAAGGTCCC CCTTGGGA CCATAGAAAT ATCAGGACAG CCCAAGCGG TGGAGACTGA ACTCGCAGCT

                    nlaIV          aciI          nlaIII          nspI          haeIII/palI          haeI          aflIII
                    sfaNI          nlaIV          aciI          nlaIII          nspI          haeIII/palI          haeI          aflIII
6101 TTTTGTGAT GCTCGTCAGG GGGCGGAGC CTATGGAAGA ACGCCAGCAA CGCGCCTTT TACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCAGATGT
AAAAACACTA CGAGCAGTCC CCCCCTCTCG GATACCTTTT TCGGCTCGTT GCGCCGAAA AATGCCAAGG ACCGGAAC GACCGAANA CGAGTGATACA

                    tfII          hinfI          fnu4HI          bbsI          pleI          hinPI          hinfI          hhaI/cfoI
                    bsrBI          aciI          fnu4HI          mcrI          hhaI/cfoI
6201 TCTTCTGCTG GTTATCCCCT GATTCGTGG ATAACCGTAT TACCGCTTT GAGTGAGCTG ATACCGCTCG CCGAGCGGA ACAGCGAGC GCAGCGAGTC
AGAAAGAGC CAATAGGGA CTAAGACACC TATTGGCATA ATGGCGAAA CTCACCTGAC TATGGCGAGC GCGCTCGCT TGCTGGCTCG CGTGGCTCAG

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FIG. 9P

```

sau3AI
mboI/ndeII[dam-]
dpmI[dam+] sau3AI          thAI
dpmII[dam-] mboI/ndeII[dam-]
    bstYI/xhoII dpmI[dam+] fnuDII/mvnI
sau3AI alwI[dam-] dpmII[dam-] bstUI
mboI/ndeII[dam-] alwI[dam-] bsh1236I
dpmI[dam+] mboII[dam-] hinPI fnu4HI
dpmII[dam-] bstYI/xhoII hhaI/cfoI bbvI
5501 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAGATCTTC TTGAGATCCT TTTTCTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC
    AAAAGCAAGG TGACTCGCAG TCTGGGGCAT CTTTCTCTAGT TTCCTAGAAG AACTCTAGGA AAAAAGAGAG CCGATTAGAC GACGAACGTT TGTTTTTTGG

sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
    acII
    nspBII
    hpaII
    mspI
    haeIII/palI
    haeI
    bslI
    rmaI
    maeI
    TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAATCTT GTAGCACCGC CTACATACCT CGCTCTGCTA ATCTGTGTAC CAGTGGCTGC TGCCAGTGGC
    AGATCACATC GGCATCAATC CCGTGGTGAA GTTCTTGAGA CATCTGGCG GATGTATGA GCGAGACGAT TAGGACAATG GTCACCGAGC ACGGTCACCG
    scrFI
    nciI
    mspI
    hpaII
    dsav
    cauII
    hinfI
    pleI
    maeIII
    hhaI/cfoI
    hinPI mcrI
    bsaWI
    hpaII
    mspI
    fnu4HI
    nspBII
    acII
    hgiAI/aspHI
    bsp1286
    bsiHKA1
    bmyI
    apaLI/snoI
    alw44I/snoI
    aluI
5801 GATAAGTGT GTCTTACCG GTTGACTCA AGACGATAGT TACCGGATAA GGCGCAGCGG TCGGGCTGAA CCGGGGGTTC GTGCACACAG CCCAGCTTGG
    CTATTACGA CAGAATGGCC CAACCTGAGT TCTGTCTATCA ATGGCCTATT CCGCGTGGCC AGCCCGACTT GCCCCCCAAG CACGTGTGTC GGTTCGAACC

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FIG. 90

hinPI
 mstI
 avIII/fspI
 bsrI
 tru9I
 mseI
 maeII hhaI/cfoI
 pspl406I
 fnu4HI
 bbvI
 sfaNI
 maeIII
 CGGATAAAGT TGCAGGACCA CTTCTGGCCT CGGCCCTTCC GGTGGCTGG
 GCTTATCTG ACCTACCTCC GCTATTCTA AGTCTCTGT GAAGACGGA GCGCGGAAGG CCGACCGACC
 mspI
 hpaII
 scrFI
 aluI nciI
 rmaI dsav
 maeI cauII
 ACTTAGCTT CCGGCAACA ATTAATAGAC TGGATGGAGG CGGATAAAGT TGCAGGACCA CTTCTGGCCT CGGCCCTTCC GGTGGCTGG
 GCTTATCTG ACCTACCTCC GCTATTCTA AGTCTCTGT GAAGACGGA GCGCGGAAGG CCGACCGACC
 mspI
 hpaII
 cfr10I
 nlaIV hphI
 gsuI/bpmI
 TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC CGGATATCAT TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA
 AAATAACGAC TATTAGACC TCGGCCACTC GCACCCAGAG CGCCATAGTA ACCTCGTGAC CCGGTCTAC CATTCGGGAG GGCATAGCAT CAATAGATGT
 ddeI
 sau3AI
 nlaIV
 mboI/ndeII(dam-) mnlI
 dpnI(dam+) hgiCI
 dpnII(dam-) banI
 maeIII
 CGACGGGAG TCAGGCAACT ATGGATGAAC GAATAGACA GATCGCTGAG ATAGTGCCT CACTGATTAA GCATTGGTAA CTGTACAGACC AAGTTTACTC
 GCTGCCCTC AGTCCGTGA TACCTACTTG CTTTATCTGT CTAGCGACTC TATCCACGGA GTGACTTAAT CGTAACCAAT GACAGTCTGG TTCAAATGAG
 rmaI sau3AI
 sau3AI hphI mboI/ndeII(dam-)
 mboI/ndeII(dam-)
 dpnI(dam+) dpnII(dam+)
 tru9I dpnII(dam-) dpnII(dam-)
 ahaIII/draI maeI alwI(dam-)
 tru9I bstVI/xhoII bstVI/xhoII
 mseI mseI alwI(dam-) mboII(dam-)
 ahaIII/draI mseI mseI alwI(dam-) mboII(dam-)
 nlaIII rcaI
 maeII
 tru9I
 mseI
 ATATATACTT TAGATTGATT TAAACTTCA TTTTAAATTT AAAAGGATCT AGGTGAAGAT CCTTTTGTAT AATCTCAIGA CCAAAATCCC TTAACGTGAG
 TATATATGAA ATCTAACTAA ATTTTGAAGT AAAAATTA TTTTCTAGA TCCACTTCTA GGAAACTA TTAGAGTACT GGTTTTAGGG AATTGCACTC

FIG. 9N

FIG. 9N

4601

hphI sfanI mboII[dam-] alw44I/snoI maeII taqI alwI[dam-] aciI bstYI/xhoII mboII
CTGGTGAAG TAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGTTA CATCGAACTG GATCTCAACA GCGTAAAGAT CCTGAGAGT TTTGGCCCCG
GACCACITTC ATTCTCTACG ACTTCTAGC AACCCACGTG CTCACCCAAT GTAGCTTGAC GTAGAGTTGT CGCCATTCTA GGAACCTCTCA AAAGCGGGG

4701

maeII hgIAI/aspHI
psp1406I bsp1286 tru9I
xmnI bsiHKAi mseI
asp700 bmyI ahaIII/draI
TTCTTGCNAA AGGTTACTAC TCGTGAANAAT TTCAAGACGA TACACCGCG CATATAGGG CACTACTGCG GCCCGTTCTC GTTGAGCCAG CGCGGTATGT

4801

ddeI csp6I bsrI
scaI hphI maeIII sfanI fokI nlaIII
CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAGCATC TTACGGATGG CATGACAGTA AGAGAAATTAT GCAGTGTCTGC CATAACCATG
GATAAGATC TTACTGAACC AACTCATGAG TGGTCAGTGT CTTTCTGAGT AATGCTTACC GTACTGTCTAT TCTCTTAATA CGTCACGACG GTATTGTATC

4901

haeIII/palI
eaeI
cfrI
fnu4HI
aciI
sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
pvuI/bspCI
mcrI mnlI
CTGCGGCCAA CTTACTTCTG ACAGACTCG GAGGACCGAA GGAGCTAACCC GCTTTTTCG ACAACATGGG GCATCATGTA ACTCGCCTTG
TCACTATTGT GAGCCCGGTT GAATGAAGAC TGTGTCTAGC CTCCTGGCTT CCTCGATTGG CGAAAAACG TGTGTACCC CCTAGTACAT TGAGCGGAAC

FIG. 9M

hinPI
 hhaI/cfoI
 thaI
 fnuDII/mvnl
 bstUI
 nspBII bsh1236I
 aciI hgaI
 nspBII bsh1236I
 aciI hgaI
 4101 ACAATCTGCT CTGATGCCG ATAGTTAAGC CAACCTCGCT ATCGCTACGT GACTGGGTCA TGGCTGGCC CGACACCCG CCAACACCCG CTGACGGCCG
 TGTTAGACGA GACTACGGG TATCAATTG GTTAGGCGA TAGCGATGCA CTGACCCAGT ACCGACGGG GGTGTGGG GGTGTGGG GACTGGCCG

hinPI
 fnu4HI
 maeIII
 nlaIII hhaI/cfoI
 bsaAI tth1111/aspI bbvI
 aciI
 4201 CTGACGGGCT TGTCTGCTCC CGGCATCCG TTACAGACAA GCTGTGACCG TCCTCGGGAG CTGCATGTGT CAGAGGTTT CACCGTCATC ACCGAAACGC
 GACTGCCCGA ACAGACGAGG GCGTAGGCG AATGTCTGTT CGACACTGGC AGAGGCCCTC GACGTACACA GTCTCCAAA GTGGCAGTAG TGGCTTTGCG

mspI
 hpaII
 scrFI
 nciI
 dsaV sfaNI
 cauII foki
 aluI maeIII
 cauII bbvI nlaIII
 mnlI hpaII fnu4HI
 bspI
 esp3I hpaII fnu4HI
 bspI
 dsaV aluI nspHI
 cauII bbvI nlaIII
 mnlI hpaII hphI
 bsh1236I
 4301 GCGAGGCAGT ATTCTTGAG ACAGAAAGGC CTCGTGATAC GCCTATTTT ATAGTTAAT GTCATGATAA TAATGGTTTC TTAGAGTCA GGTGGCATT
 CGCTCCGTC TAAGAACTTC TGCTTTCCCG GAGCACTATG CGGATAAAAA TATCCAATTA CAGTACTATT ATTACCAAAG AATCTGCAGT CCACCGTGAA

mnlI
 haeIII/palI
 sau96I
 asuI
 eco0109I/draII
 maeII
 hinII/acyI
 ahaII/bsaHI
 ddeI aatII

nlaIV
 aciI
 thaI
 fnuDII/mvnl
 bstUI
 bsh1236I
 hinPI
 hhaI/cfoI
 4401 TTCGGGGAAA TGTGGCGGA ACCCCTATT GTTTATTTT CTAAATACAT TCAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA
 AAGCCCCCTT ACACGGCCT TGGGGATAA CAATAAAAA GATTTATGTA AGTTTATACA TAGGCGAGTA CTCTGTTATT GGGACTATTT ACGAAGTTAT

mboII
 earI/ksp632I
 4501 ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTCCGT GTCGCCCTTA TTCCCTTTT TCGGGCATTT TGCCTTCCGT TTTTGTCTCA CCCAGAAACG
 TATAACTTTT TCCTTCTCAT ACTCATAAGT TGTAAAGGCA CAGCGGGAAT AAGGAAAAA ACGCCGTAAA ACAGAAAGGAC AAAAACGAGT GGGTCTTTGC

mboII
 earI/ksp632I
 4601 ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTCCGT GTCGCCCTTA TTCCCTTTT TCGGGCATTT TGCCTTCCGT TTTTGTCTCA CCCAGAAACG
 TATAACTTTT TCCTTCTCAT ACTCATAAGT TGTAAAGGCA CAGCGGGAAT AAGGAAAAA ACGCCGTAAA ACAGAAAGGAC AAAAACGAGT GGGTCTTTGC

FIG. 9L

hinPI fnu4HI acilI fnu4HI hinPI fnu4HI
 hhai/cfoI acilI hhai/cfoI hhai/cfoI
 thalI fnuDII/mvnI thalI fnuDII/mvnI
 bstUI bstUI bstUI
 bsh1236I bsh1236I bsh1236I
 rsaI scfI fnu4HI tru9I bsh1236I maeII bsh1236I acilI hinPI
 csp6I bsh1236I maeII hhai/cfoI maeII bsh1236I hhai/cfoI
 3601 CACCGCATAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGCGCATTA AGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG
 GTGGCGTATG CAGTTTCGTT GGTATCATGC GCGGACATC GCGCGTAT TCGCGCGCC CACACCACCA ATGCGCGTCG CACTGCGCAT GTGAACGGTC
 hinPI nlaIV
 hhai/cfoI hgiJII
 haeII bsp1286
 rnaI bsrBI bmyI
 maeI acilI maeII banII nlaIV
 3701 CGCCCTAGCG CCGCTCCTT TCGTTCTT CCGTCTCTT CCGCTCCTT TCGCGGCTT TCGCGGCTT TCGCGGCTT TCGCGGCTT TCGCGGCTT TCGCGGCTT
 GCGGATCGC GCGGAGGAA AGCGAGGAA GCGGAGGAA GCGGAGGAA GCGGAGGAA GCGGAGGAA GCGGAGGAA GCGGAGGAA GCGGAGGAA GCGGAGGAA
 nlaIV haeII haeII/pali
 hgiCI taqI draIII sau96I maeII
 bani mnlI hphI bsaI asuI drdI
 3801 CGATTAGTG CTTACGGCA CCTCGACCC AAAAAGCTT ATTTGGTGA TGGTTCACGT AGTGGGCGCAT CGCCCTGATA GACGGTTTTT CGCCCTTTGA
 GCTAATCAC GAATGCCGT GGAGCTGGG TTTTGTGAAC TAAACCACT ACCAAGTGA TCACCGCGTA CCGGCGTAT CTGCCAAAA GCGGGAACCT
 pleI tru9I pleI bslI
 hinPI maeII hinPI bsrI bslI auaI
 3901 CGTTGGAGTC CAGTTCTTT AATAGTGGAC TCTGTGTCCA AACTGGAACA ACACCAACC CTATCTGGG CTATCTCTTT GATTATAAG GGATTTTGGC
 GCAACCTCAG GTCAAGAAA TTATCACCTG AGAACAAAGT TTGACCTTGT TGTGAGTTGG GATAGAGCCC GATAAGAAAA CTAAATATTC CCTAAAACGG
 tru9I tru9I tru9I apoI tru9I maeII
 maeI maeI maeI apoI tru9I maeII
 bsh1236I bsh1236I bsh1236I
 4001 GATTTCGCC TATTGGTTAA AAAATGAGCT GATTAAACAA AAATTTAAG CGAATTTTAA CAAATATTA AGTTTACAA TTTTATGGTG CACTCTCAGT
 CTAAGCGG ATAACCAATT TTTTACTCGA CTAATTTGTT TTTTATTAAT TCGAATGTT AAAATACCAC GTGAGAGTCA

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FIG. 9K

[illegible]

FIG. 9J

```

          taqI(dam-)
          claI/bsp106(dam-)
          sau3AI
          mboI/ndeII(dam-)
          dpnII(dam+)
          dpnII(dam-)
          nlaIII aluI(dam-)
2801 AAAGCAATAG CATCAAAAT TTCACAAATA AAGCAATTTT TTCACTGCAT TCTAGTTG GTTGTCCAA ACTCATCAAT GTATCTTATC ATGCTGGAT
      TTTCTGTTATC GTAGTGTTTA AAGTGTTTAT TTCGTAATAA AAGTGACGTA AGATCAACAC CAACAGGTT TGAGTAGTTA CATAGNATAG TACAGACCTA
          rmaI
          bsmI maeI
          rsal
          csp6I
          nlaIV
          kpnI
          hgiCI
          banI
          asp718 mnlI
          acc65I ddeI aciI
          pvuII
          nspBII
2901 CGATCGGAA TTAATTGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACCT GGTAGGTAC CTTCTAGGC GGAAGAAC AGCTGTGAA
      GCTAGCCCTT AATTAAGCCG CGTGTGGTA CCGGACTTTA TTGGAGACTT TCTCCTGAA CCAATCCATG GAAGACTCCG CTTTCTTGG TCGACACCTT
          nlaIV
          scrFI
          mvaI
          ecorII
          dsav
          bstNI
          apyI(dcm+)
          bsaJI
          ppul01
          nsII/avaIII
          nlaIII
          sphI
          nspI sfanI
          nspHI
          aciI
          foki
          aciI
3001 TGTGTGTCAG TTAGGGTGTG GAAAGTCCC AGGTCGCCA GCAGGCAGAA GTATGCAAG CATGCATCTC AATTAGTCAG CAACGAGTG TGGAAAGTCC
      ACACACAGTC AATCCACAC CTTTCAGGG TCCGAGGGT CGTCCGTCTT CATACGTTTC GTACGTAGAG TTAATCAGTC GTTGCTCCAC ACCTTTCAGG
          nlaIV
          scrFI
          mvaI
          ecorII
          dsav
          bstNI
          apyI(dcm+)
          bsaJI
          ppul01
          nsII/avaIII
          nlaIII
          sphI
          nspI
          nspHI
          aciI
          foki
          aciI
3101 CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATTA GTCCGCCCC TAACTCCGCC CATCCGCCCC CTAACTCCGC
      GGTCCGAGGG GTCTGCCGTC TTCATACGTT TCGTACGTAG AGTTAATCAG TCGTTGGTAT CAGGCGGGG ATTGAGGCGG GTAGGCGGG GATTGAGGCG
          nlaIV
          scrFI
          mvaI
          ecorII
          dsav
          bstNI
          apyI(dcm+)
          bsaJI
          ppul01
          nsII/avaIII
          nlaIII
          sphI
          nspI
          nspHI
          aciI
          foki
          aciI

```

FIG. 91

SUBSTITUTE SHEET (RULE 26)

FIG. 9H

FIG. 9H

```
sau96I
nlaIV
avaII
mspI
scrFI
ncil
sau3AI hpaII
mboI/ndeII(dam-)
dpnI(dam+)
nlaIII dsav
rcal
bspHI(dam-) asuI
mnlI dpnII(dam-) bsu36I/mstII/sauI
styI
bsaJI
2101 TTCCCCCAA AACCCAAAGG CACCTTCATG ATCTCCCGG CCGCTGAGGT CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCTCGAG GTCAAGTTCA
AAGGGGGTT TTGGGTTCTT GTGGGAGTAC TAGAGGGCCT GGGGACTCCA GTGTACGCAC CACCACCTGC ACTCGGTGCT TCTGGGACTC CAGTTCAAGT
acil
thai
fnuDII/mvnI
bstUI
bsh1236I
sacII/sstII
nspBII
kspl
dsaI
bsaJI
acil
rsal
csp6I
fnu4HI mnlI csp6I bsaAI
maeII
2201 bsrI bsaAI mnlI rsal csp6I maeII bsaAI hgaI hphI bslI
GGAGCGGTG GAGGTGCATA ATGCCAAGAC AAAGCCGCGG GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTACCGTTC TCACGTCTCT
TGACCATGCA CCTGCCGCAC CTCCACGTAT TAGGGTTCTG TTTCCGGCGC CTCTCTGCTCA TGTGTGCTG CATGGCACAC CAGTCGCAGG AGTGGCAGGA
maeII
rsal
csp6I
bsrI bsaAI mnlI
2201 mval bsrI
ecoRII
dsav
bstNI
apyl(dcm+)
2301 GCACCAAGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA AAGCCCTCCC AGCCCCCATC GAGAAACCA TCTCCAAAGC CAAAGGCGAG
CGTGGTCTTG ACCGACTTAC CGTTCTCTCAT GTTCACGTTTC CAGAGTTGT TTCGGGAGGG TCCGGGGTAG CTCCTTTGGT AGAGTTTTCG GTTTCCCGTC
fnu4HI
bbvI
```


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FIG. 9G

```

scrFI      hinPI      nlaIV
mvaI      narI      kasi
ecorIII   hinII/acyI hgiCI
ecoNI     hphI      haeII
dsav      mspI      bspI286
bstNI     hpaII     bsiHKA I
bsLI      cfrI01    bmyI
apyI(dcm+) bsaWI tthI111I/aspI fnu4HI
fnu4HI     bslI ageI maeIII ddeI hhaI/cfoI nspBII aciI apaLI/snoI dsav
bbvI      bsu36I/mstII/sauI ddeI hhaI/cfoI nspBII alw44I/snoI cauII scfI
1801 GGTGCTGG TCAAGGACTA CTTCCCGGAA CCGGTGACGG TGTCGTGGAA CTCAGGGGCC CTGACCAGCG GCGTGCACAC CTTCCCGGCT GTCTACAGT
CCGACGGACC AGTTCTGTAT GAAGGGGCTT GGCACCTGCC ACAGCACCTT GAGTCCGGCG GACTGGTGG CCGACGTGGT GAAGGGCCGA CAGGATGTCA

fnu4HI     ddeI pleI      nlaIV
mnlI hinfi      fnu4HI hgiCI
eco8II mnlI bbvI      bspI286
bsu36I/mstII/sauI ddeI hhaI/cfoI nspBII alw44I/snoI cauII scfI
1901 CTTACAGGACT CTACTCCCTC AGCAGGTGG TGACTGTGG CTTCTAGCAGC TTGGGCACCC AGACCTACAT CTGCAACGTG ANTACAAGC CCAGCAACAC
GGAGTCCTGA GATGAGGGAG TCGTCCGACC ACTGACACCG GAGATCGTGG AACCCGTGG TCTGGATGTA GACGTGGCAG TTAGTGTTCG GTTCGTTCGTG

hgiJII     nlaIII
bspI286    nspI
bmyI       nspHI
banII      maeIII
2001 CAAGGTGGAC AAGAAAGTTG AGCCCAAAATC TTGTGACAAA ACTCACACAT GCCCACCCTG CCCAGCACCT GAATCTCTGG GGGGACCCTC AGTCTTCCTC
GTTCACCTG TTCTTTCAAC TCGGGTTTAG AACACTGTTT TGAGTGTGTA CCGGTGGCAG GGGTCTGTGA CTTGAGGACC CCCCTGGCAG TCAGAAAGGAG

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SUBSTITUTE SHEET (RULE 26)

FIG. 9E

FIG. 9E

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FIG. 9D

[illegible]

FIG. 9C

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```

      haeIII/palI
      haeI
scrFI      scrFI      mvaI      mvaI      mvaI      mvaI      mvaI      mvaI      mvaI      mvaI
      ecorII      ecorII      ecorII      ecorII      ecorII      ecorII      ecorII      ecorII
dsav      tfil      dsav      bstNI      nlaIII      bstNI      ddel      pleI      binfi
      apyI(dcm+)      hinfi      apyI(dcm+)      apyI(dcm+)      apyI(dcm+)      apyI(dcm+)      apyI(dcm+)      apyI(dcm+)
801 ACAACCGAA TTGCAAGTA AGTAGACAT GGTTTGATA GTCCGAGGCA GTTCTGTTTA CCAGGAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT
TGTGGCCTT AACCGTTCTT TCAATCTGTA CCAACCTAT CAGCCTCCGT CAAGACAAAT GGTCTCTGGG TACTTAGTGG GTCCGGTGGG ATCTGAGAAA

      accI      nlaIII      mnlI
      mspI      hpaII      bsaBI
      mboI/ndeII(dam-)
      dpnI(dam+)
      dpnII(dam-)
      maeIII      alwI(dam-)      apoI      maeIII      mnlI      bsaJI      bslI      ddel
901 GTGACAGGA TCATCCAGGA ATTTGAAAGT GACACGTTTT TCCAGAAAT TGATTTGGG AAATATAAAC CTCTCCAGA ATACCAGGC GTCTCTCTG
CACTGTTCT AGTACGCTCT TAACTTTCA CTGTGCNAAA AGGTCCTTTA ACTAAACCCC TTATATTTG GAGAGGCTCT TATGGGTCCG CAGGAGAGAC

      scrFI      mvaI      ecorII      dsav      bstNI      apyI(dcm+)      sau96I      avall
      mnlI      sfaNI      mboII      mboII      mnlI      aluI
1001 AGGTCCAGGA GGAAGAAGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG AAAGACTAAC AGGAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAGCT
TCCAGGCTCT CCTTTTCCG TAGTTCATAT TCAAACTTCA GATGCTCTTC TTTCGTATG TCCTTCTACG AAAGTTCAAG AGACGAGGG AGGANTTCGA

      styI      bsaJI
      sau3AI
      mboI/ndeII(dam-)
      dpnI(dam+)
      dpnII(dam-)
      alwI(dam-)
      bstYI/xhoII
      ppu101      nsii/avaIII      bsaJI
      aluI      tru9I      mseI      asel/asnI/vspI
1101 ATGCAATTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCTTGG CTTCGTTAGA ACCGAGCTAC AATTAATACA TAACCTTATG TATCATACAC
TAGGTAAAAA TATTCTGGTA CCTGAAAC GACCGAATC TAGGGGAACC GAAGCAATCT TCGCTCGATG TTAATTATGT ATTGGAATAC ATAGTATGTG

```

BNSDOCID: <WO___9604391A1_I_>

FIG. 9A

FIG. 9A

```

aluI          sau3AI pvuII          nlaIV
sstI          mboI/ndeII[dam-]      scrFI
sacI          dpnI[dam+]            mvaI
hgiAII       pvuI/bspCI             ecoRII
hgiAI/aspHI  pleI dpnII[dam-]         dsav
ecII36II     hinfi taqI[dam-]        bstNI
bsp1286      rmaI mcrI nspBII        apyI[dcm+]
bsiHKA1      maeI taqI[dam-]         bsaJI
bmyI         rnaI maeI taqI[dam-]    nlaIV
banII        taqI                    sfanI
                                     ppulOI
                                     nsiI/avaIII
                                     nlaIII
                                     sphI
                                     nspI
                                     nspHI
101 GAAGTATGCA AGCATGTCAT CTCAAATTAGT CAGCAACCGAG GTGTGGAAAG TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA
CTTCATACGT TTCGTACGTA GAGTTAATCA GTCGTTGGTC CACACCTTTC AGGGGTCCGA GGGGTCCGTC GTCTTCATAC GTTTCGTACG TAGAGTTAAT

                                     nlaIII
                                     styI
                                     ncoI
                                     bslI dsal
                                     acil bsaJI
201 GTCAGCAACC ATAGTCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC CGCCAGTTC CGCCCATTTCT CGCCCCCATG GCTGACTAAT TTTTTTATT
CAGTCGTTGG TATCAGGCG GGGATTGAG GGGTAGGGC GGGGATTGAG GCGGTCAG GCGGTCAG GCGGGGTAC GCGCTGATTA AAAAAATAA

                                     rmaI
                                     styI
                                     bsaJI
                                     blnI
                                     avrII
                                     haeIII/palI
                                     stuI
                                     haeI
                                     mnII maeI
                                     mnII
                                     mnII
                                     haeIII/palI
                                     haeIII/palI
                                     mcrI
                                     eagI/xmaIII/ecII
301 TATGACAGG CCGAGGCGC CTCGGCTCT GAGCTATTCC AGAAGTAGTG AGGAGGCTTT TTGGAGGCC TAGGCTTTTG CAAAAGCTA GCTTATCCG
ATACGTCTCC GGCTCCGGC GAGCCGGAGA CTCGATAAGG TCTTCATCAC TCCTCCGAAA AAACCTCCCG ATCCGAAAC GTTTTCGAT CGAATAGGC

```

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FIG. 8

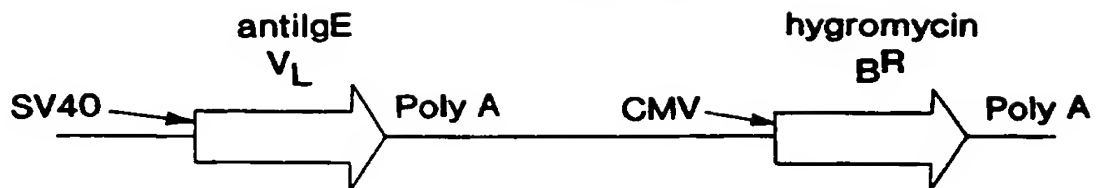
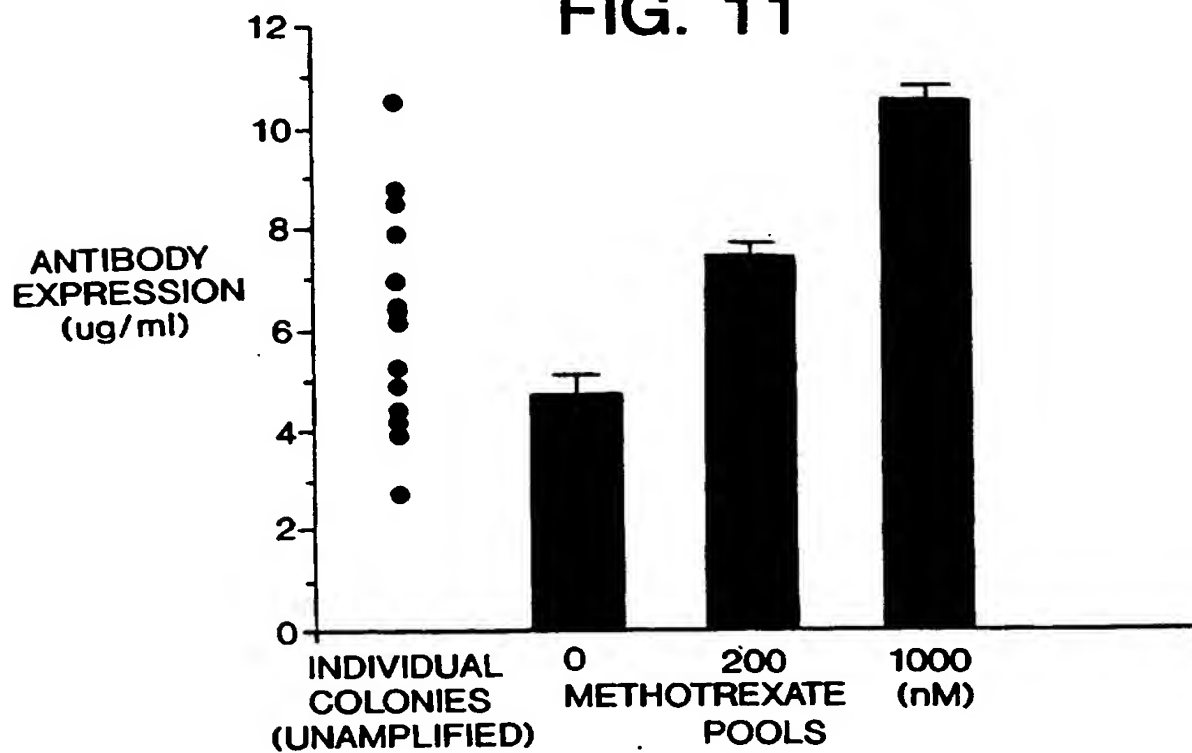


FIG. 11



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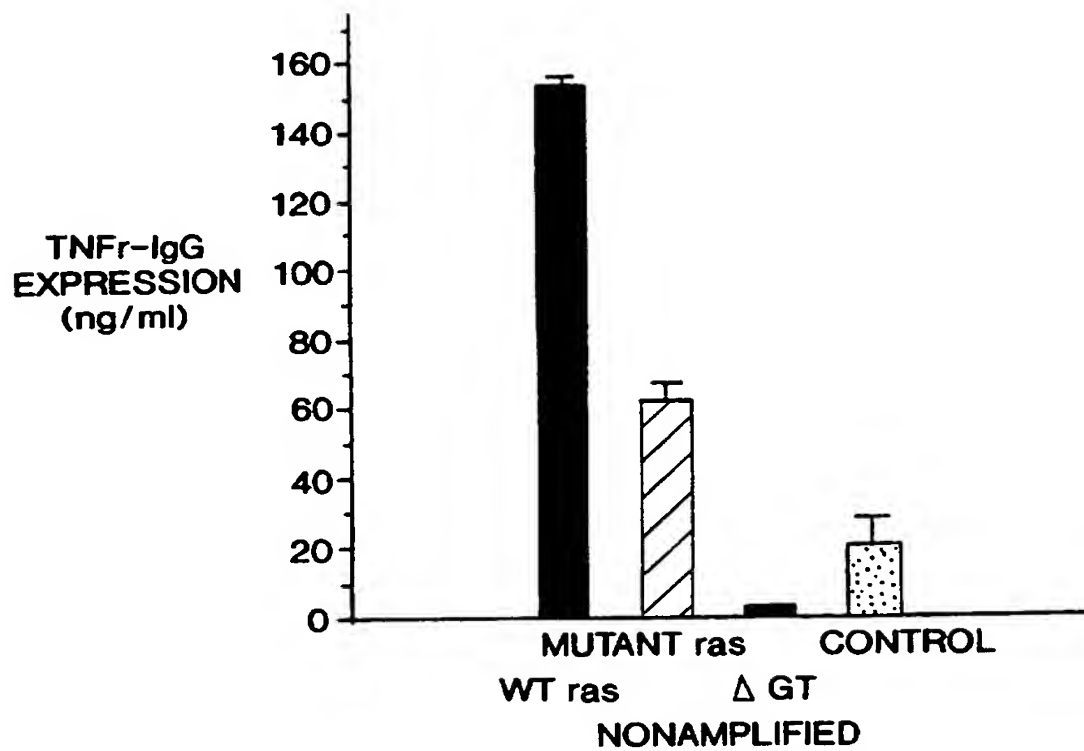


FIG. 7A

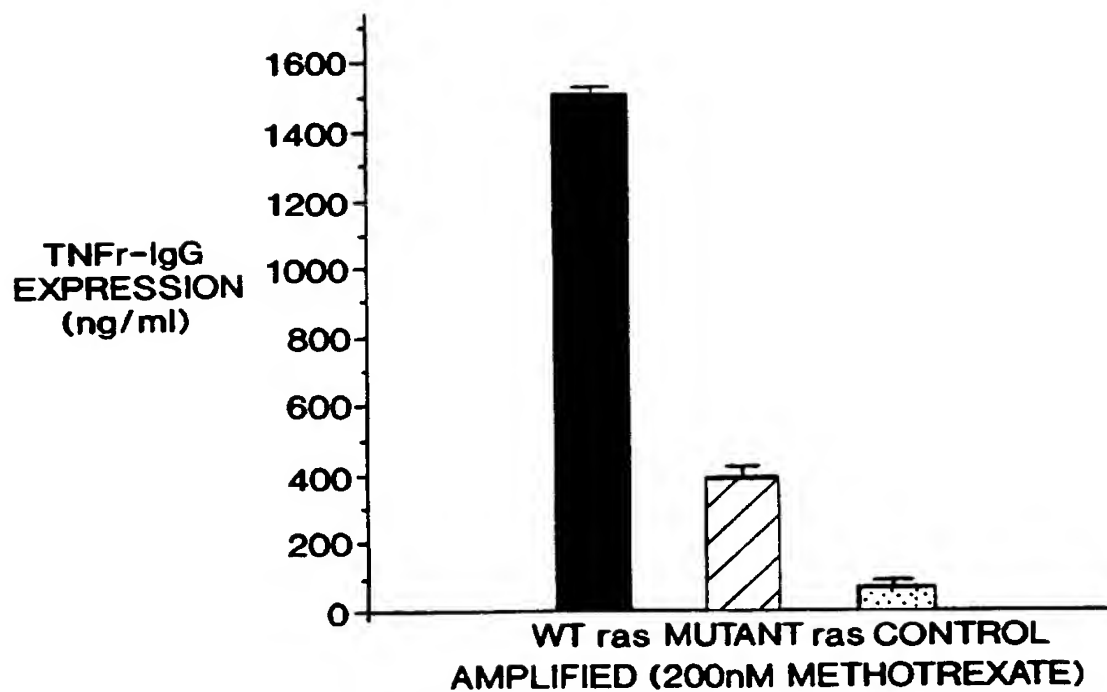


FIG. 7B

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FIG. 6R

```

haeIII/palI
haeI
scrFI
mvaI bslI
ecorII
dsav
nlaIII
nspi
haeIII/palI nspHI
bstNI
apyI[dcM+]
haeI
haeIII/palI afillI
6501 CCTGGCCCTT TGCTGCACAT GTTCTTTCTT GGTATATCCC CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT
GGACCCGGAA ACACCGGAA AACGAGTGTA CAAGAAGGA CGCAATAGG GACTAAGACA CCTATTGGCA TAATGGCGGA AACTCACTCG ACTATGGCGA
bsrBI
aciI
aluI
6601 CGCCGACGCC GAACGACCGA CGCAGCGGAG TCAGTGAGCG AGGAAGCGGA AGAGCGCCCA ATACGCAAC CGCTCTCCC CGCGGTGG CCGATTCAAT
CGCGGCTCGG CTTGCTGGCT CGCGTGGCT AGTCACTGCG TCCTTGCCTT TCTGCGGGT TATGCGTTG GCGGAGAGGG GCGCGCAACC GGCTAAGTAA
thaI
fnuDII/mvni
bstUI
bsh1236I
hinPI
hhaI/cfoI
thaI
fnuDII/mvni
bstUI haeIII/palI
bsh1236I
tru9I
bslI eaeI tfil ael/asnI/vspI
aciI cfrI hinFI mseI
6601 CGCGGCTCGG CTTGCTGGCT CGCGTGGCT AGTCACTGCG TCCTTGCCTT TCTGCGGGT TATGCGTTG GCGGAGAGGG GCGCGCAACC GGCTAAGTAA
scrFI
mvaI
ecorII
dsav
nlaIV bstNI
hgiCI apyI[dcM+]
banI bsaJI
6701 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC CAGTCACTCAT CCTCACTCAT TAGGCACCCC AGGCTTTACA
TTAGGTCGAC CGTGCTGTCC AAAGGGCTGA CCTTTCGCC CCTTTCGCC GTCACTGCGG TCGCTTAAT TACACTCAAT GGAGTGAGTA ATCCGTGGG TCCGAATGT
tru9I
mseI
aseI/asnI/vspI
xmnI
asp700
6801 CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT TTCACACAGG AACAGCTAT GACCATGATT ACGAATTAA
GAAATACGAA GGCCGAGCAT ACAACACACC TTAACACTCG CCTATTGTTA AGTGTGTCC TTTGTGATA CTGGTACTAA TGCTTAATT

```

>length: 6889

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FIG. 6Q

```

        scrFI      aciI      nspBII      fnu4HI      bbvI      hinPI      mcrI      hhaI/cfoI
        nciI      mspI      hpaII      bsaWI      maeIII      pleI      hinFI      cauII      hinfI
        dsav      ggtTGGGCT  GCGATAAGTC  GGTCTTTACC  GGTGTGGACT  CAAGACGATA  GTTCTGCTAT  CAATGGCCTA  TTCCGGCTCG  CCAGCCCGAC
        6101  TAATCCTGTT  ACCAGTGGCT  GCTGCCAGTG  GCGATAAGTC  GGTCTTTACC  GGTGTGGACT  CAAGACGATA  GTTCTGCTAT  CAATGGCCTA  TTCCGGCTCG  CCAGCCCGAC
        ATTAGGACAA  TGGTCACCGA  CGACGGTCAC  CGCTATTTCG  CACAGAATGG  CCAACCTGA  GTTCTGCTAT  CAATGGCCTA  TTCCGGCTCG  CCAGCCCGAC

        hgiAI/aspHI  bsp1286  bsiHKA1  bmyI  apaLI/snoI  alw44I/snoI  aluI  ddeI      scfI      hinPI      hhaI/cfoI      haeII
        6201  AACGGGGGCT  TCGTGCACAC  AGCCAGCTT  GGAGCGAAGC  ACCTACACCG  AACTGAGATA  CCTACAGCGT  GAGCATTGAG  AAAGGCCAC  GCTTCCCGAA
        TTGCCCCCA  AGCAGCTG  TCGGTCGAA  CCTCGCTGC  TGGATGTGGC  TTGACTCTAT  GGATGTGCA  CTCGTAAC  CTCGTAAC  TTCGGCGTG  CGAAGGGCTT

        scrFI      mvaI      ecorII      dsav      bstNI      bsaJI      hinPI      mnII      hhaI/cfoI      aluI      apyI[dcn+]      apyI[dcn+]
        6301  GGGAGAAAG  CGGACAGGTA  TCCGGTAAGC  GGCAGGGTCG  GAACAGGAGA  GCGCAGGAGG  GAGCTTCCAG  GGGGAAACGC  CTGTATCTT  TATAGTCCG
        CCTCTTTCC  GCCTGTCCAT  AGGCCATTCC  CCGTCCACG  CTTGTCTCT  CCGTGTCTCC  CTCGAAGGTC  CCCCTTTGCG  GACCATAGAA  ATATCAGGAC

        haeIII/palI  fnu4HI      aciI      thaI      bslI      fnuDII/mvnI      bstUI      bsh1236I      nlaIV
        6401  TCGGGTTTCG  CCACCTCTGA  CTTGAGCGTC  GATTTTTCG  ATGCTCGTCA  GGGGGCGGGA  GCCTATGGAA  AAACGCCAGC  AACGGCGCT  TTTTACGGTT
        AGCCCAAAGC  GGTGGAGACT  GAACTCGCAG  CTAAAAACAC  TACGAGCAGT  CCCCCGCT  CGATACCTT  TTTGCGGTG  TTGCGCCGGA  AAAATGCCAA

```

FIG. 6P

[illegible]

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FIG. 60

```

sau96I
avail
sau3AI asuI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
pvuI/bspCI
mcrI mnlI aluI aciI
haeIII/palI
eaeI
cfrI
fnu4HI
bbvI nlaIII
5201 TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTT
ATTCTCTTAA TAGCTACGA CCGTATTGGT ACTCACTATT GTGACGCCGG TTGAATGAAG ACTGTTGCTA GCCTCCTGSC TTCTCTGATT GGCGAAAAAA

fnu4HI
bbvI nlaIII
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
fnu4HI
bbvI nlaIII
5301 GCACAACATG GGGGATCATG TAACTGCGCT TAACCTGCGT GAGGCGGAGC TGAATGAAGC CATAACCAAC GACGAGCGTG ACACCAAGAT GCCAGCAGCA
CGTGTGTGAC CCCTAGTAC ATTGAGCGGA ACTAGCAACC CTTGGCCTCG ACTTACTTCG GTATGGTTTG CTGCTCGCAC TGCTGCTGCTA CGCTCGTCTG

hinPI
mstI
aviII/fspI bsrI
maeII hhaI/cfoI tru9I mseI
psp1406I
5401 ATGGCAACAA CGTTGCGCAA ACTATTAACT GCGGAACCTAC TTACTCTAGC TTCCCGGCCAA CAATTATAG ACTGGATGGA GGCGGATAAA GTTGACGAGC
TACCGTTGTT GCAACGCGTT TGATAATTGA CCGCTTGATG AATGAGATCG AAGGCGCGTT GTTAATTATC TGACCTACTT CCGCTTATTT CAACGCTCCTG

hinPI
hpaII
scrFI
aluI nciI
rmaI dsav
maeI cauII
5501 CACTTCTGCG CTCGCGCCCTT CCGGCTGGCT GGTATTATTC TGATAAATCT GGAGCCGGTG AGCGTGGGTC TCAGCGGATC ATTGACGAC ATTGCGCCAGA
GTGAAGACGC GAGCCGGGAA GGCCGACCGA CCAATAACG ACTATTTAGA CCTCGGCCAC CTCGCGCCAC TCGCACCCAG AGCGCCATAG TAACGCTGCTG ACCCGGCTCT

bglI
sau96I
haeIII/palI
hinPI asuI mspI
hhaI/cfoI hpaII
5601 TGSTAAAGCCC TCCTGATATCG TAGTTATCTA CACGACGGGG ACTCAGGCAA CTATGGATGA ACAGAAATAGA CAGATCGCTG AGATAGTGC CTCACGTGATT
ACCATTCGGG AGGCGATAGC ATCAATAGAT GTGCTGCCCC TCAGTCCGTT GATACCTACT TGCTTTATCT GTCTAGCGAC TCTATCCAG GAGTGACTAA

pleI
hinFI
eam1105I
fokI
5601 TGSTAAAGCCC TCCTGATATCG TAGTTATCTA CACGACGGGG ACTCAGGCAA CTATGGATGA ACAGAAATAGA CAGATCGCTG AGATAGTGC CTCACGTGATT
ACCATTCGGG AGGCGATAGC ATCAATAGAT GTGCTGCCCC TCAGTCCGTT GATACCTACT TGCTTTATCT GTCTAGCGAC TCTATCCAG GAGTGACTAA

```

FIG. 6N

4701 AATAATGTT TCCTAGACGT CAGGTGGCAG TTTTCGGGGA AATGTGGCGG GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAATAT GTATCCGCTC
 TTAATTACAA AGAATCTGCA GTCCACCGTG AAAAGCCCTT TTACACCGCG CTTCGGGATA AACCAATAAA AAGATTATG TAAGTTTATA CATAGGCGAG
 hinII/acyI nlaIV
 ahaII/bsaHI aciI
 aatII thal
 ddeI maeII fnuDII/mvnI
 bsmAI bstUI bsh1236I
 hinPI hhaI/cfoI
 mboII mboII
 earI/ksp632I
 sspI
 4801 ATGAGACAAT AACCCGTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG TATGAGTATT CAACATTTCC GTGTGCGCCT TATTCCTTTT TTTGCGGCAT
 TACTCTGTTA TTGGGACTAT TTACGAAGTT ATTATAACTT TTTCCTTCTC ATACTCATAA GTTGTAAGG CACAGCGGA ATAAGGGAAA AACGCCGTA
 hgiAI/aspHI
 bsp1286
 sau3AI bsiHKA1
 mboI/ndeII(dam-) bmyI
 dpnII(dam-) dpnII(dam-) apaLI/snoI
 eco57I
 hphI hphI hphI
 4901 TTTGCCCTCC TGTTTGTCT CACCCAGAAA CGCTGGTGAA AGTAAAGAT GCTGAAGATC AGTGGGTGC ACAGTGGGT TACATCGAAC TGGATCTCAA
 AAACGGAAGG ACAAAAACGA GTGGGTCTTT GCGACCACTT TCATTTTCTA CGACTTCTAG TCAACCCACG TGCTCACCCA ATGTAGCTTG ACCTAGAGTT
 mboI/ndeII(dam-) maeIII taqI alwI(dam-)
 sau3AI mboI/ndeII(dam-) dpnI(dam+) bstyI/xhoII
 bsrI dpnII(dam-)
 aciI
 thal
 fnuDII/mvnI
 bstUI
 bsh1236I
 hinPI
 hhaI/cfoI
 5001 CAGCGTAAG ATCCTTGAGA GTTTTCGCC CGAAGAACGT TTTCCAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC
 GTCGCCATTC TAGGAACCTCT CAAAAGCGGG GCTTCTTGCA AAAGGTACT ACTCGTGANA ATTCAAGAG GATACACCGC GCCATAATAG GGCACACTCTG
 nspBII bstyI/xhoII
 hhaI/cfoI
 ahaII/bsaHI
 scrFI
 nciI
 mspI
 hpaII
 dsav
 5101 GCCGGGCAAG AGCAACTCGG TCGCCGCGATA CACTATTCTC AGAATGACTT GGTGAGTAC TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG
 CGGCCCGTTC TCGTTGAGCC AGCGGCGGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGGTCACT GTCTTTTCGT AGAATGCCTA CCGTACTGTC
 rcaI
 bspHI
 bsrBI
 aciI nlaIII
 rsaI
 csp6I bsrI
 scaI hphI maeIII
 sfaNI foki nlaIII

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FIG. 6M

4201	ATCGCCCTGA	TAGACGGTTT	TTCGCCCTTT	GACGTTGGAG	TCCACGTTCT	TTAATAGTGG	ACTCTGTTTC	CAAACTGGAA	CAACACTCAA	CCCTATCTCG
	TAGCGGGACT	ATCTGCCAAA	AAGCGGGAAA	CTGCAACCTC	AGGTGCAAGA	AATTATCACC	TGAGAACAAAG	GTTTGACCTT	GTTGTGAGTT	GGGATAGACC

maeII pleI tru9I pleI bsrI bsrI bsrI

drdI hinfI maeII mseI hinfI

[illegible][illegible][illegible][illegible]

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FIG. 6L

hinPI
 hhaI/cfoI
 nlaIV
 nari
 kasi
 hinII/acyI
 hgiCI
 haeII
 bani
 sfaNI
 ahaII/bsaHI
 bglI
 acII
 fnu4HI
 acII
 thal
 fnuDII/mvni
 bstUI
 hinPI
 hhaI/cfoI
 hinPI
 thal
 fnuDII/mvni
 bstUI
 scfi
 bsh1236I
 rsaI
 hhaI/cfoI
 fnu4HI
 tru9I
 bsh1236I
 csp6I
 bslI
 acII
 mseI
 hhaI/cfoI
 mspI
 hpaII
 naeI
 maeII
 cfr10I
 maeII
 haeIII/palI
 draII
 sau96I
 bsaI
 asuI
 hphI
 haeII
 sau96I
 sau3AI
 mboI/ndeII[dam-]
 sau96I
 haeIII/palI
 asuI
 mnlI
 dpnI[dam+]
 dpnII[dam-]
 pvuI/bspCI
 merI
 mboII
 aciI
 pvuI/bspCI
 earI/ksp632I
 merI
 aluI
 pvuII
 nspBII
 foki
 CATCCCCCT TCGCCAGCTG GCGTAATAGC GCGAGGCGCC GCACCGATCG CCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGGC CTGATGCGGT GTAGGGGGA AGCGTCGAC CGCATTATCG CTCTCCGG CGTGGTAGC GGAAGGGTT GTCAACGCAT CGGACTTACC GCTTACCOCG GACTACGCCA
 3801
 fnu4HI
 hinPI
 hhaI/cfoI
 thal
 fnuDII/mvni
 bstUI
 sfaNI
 aciI
 maeII
 acII
 maeII
 csp6I
 bslI
 acII
 rsaI
 hhaI/cfoI
 fnu4HI
 tru9I
 bsh1236I
 csp6I
 bslI
 acII
 mseI
 hhaI/cfoI
 mspI
 hpaII
 naeI
 maeII
 cfr10I
 maeII
 haeIII/palI
 draII
 sau96I
 bsaI
 asuI
 hphI
 haeII
 sau96I
 sau3AI
 mboI/ndeII[dam-]
 sau96I
 haeIII/palI
 asuI
 mnlI
 dpnI[dam+]
 dpnII[dam-]
 pvuI/bspCI
 merI
 mboII
 aciI
 pvuI/bspCI
 earI/ksp632I
 merI
 aluI
 pvuII
 nspBII
 foki
 CATCCCCCT TCGCCAGCTG GCGTAATAGC GCGAGGCGCC GCACCGATCG CCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGGC CTGATGCGGT GTAGGGGGA AGCGTCGAC CGCATTATCG CTCTCCGG CGTGGTAGC GGAAGGGTT GTCAACGCAT CGGACTTACC GCTTACCOCG GACTACGCCA
 3901
 ATTCTCTCT TACGCATCTG TCGGGTATT CACACCGCAT ACGTCAAGC AACCATAGTA CGGCCCTGT AGCGCGCAT TAAGCGCGC GGGTGTGGT TAAAGAGGA ATCGGTAGC ACGCCATAA GTGTGGCGTA TGAGTTTCG TTGTATCAT CGCGGGACA TCGCGCGTA ATTGCGCGC CCCACACCAC
 4001
 GTTACGGCA GCGTGACCG TACACTTGC AGCGCCTAG CGCCGCTCC TTTCGCTTC TTCTCGCCAC GTTCGCGGC TTCTCCCGTC CAATGCGCT CGCACTGCG ATGTGAACGG TCGCGGATC GCGCGGAGG AAGCGAAAG AAGGGAAGGA AAGAGCGTG CAAGCGCGC AAAGGGGAG
 nlaIV
 hgiJII
 bsp1286
 bmyI
 baniI
 aluI
 AAGCTCTAAA TCGGGGGTCC CTTTAGGGT TCCGATTAG TCGTTACGG CACCTCGACC CCAAAAACCT TGAITGGGT GATGTTTAC GTAGTGGCC TCGAGATT AGCCCCCGAG GGAATCCCA AGGCTAAATC ACGAATGCC GTGGAGCTGG GGTTTTGA ACTAAACCA CTACCAAGT CATCACCOCG
 4101
 nlaIV
 hgiJII
 bsp1286
 bmyI
 baniI
 aluI
 AAGCTCTAAA TCGGGGGTCC CTTTAGGGT TCCGATTAG TCGTTACGG CACCTCGACC CCAAAAACCT TGAITGGGT GATGTTTAC GTAGTGGCC TCGAGATT AGCCCCCGAG GGAATCCCA AGGCTAAATC ACGAATGCC GTGGAGCTGG GGTTTTGA ACTAAACCA CTACCAAGT CATCACCOCG

FIG. 6K

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FIG. 6J

```

rmaI
mnlI
sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
nlaIV maeI hincII/hindII
bstYI/xhoII accI pStI
bamHI xbaI pleI bspMI
alwI(dam-) hinfI bspMI
3101 GGGGATCCTC TAGATCGAC CTGCAGAGC TTGGCCGCCA TGGCCCAACT TGTATTATGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCACAAT
CCCCTAGGAG ATCTCAGCTG GACGCTCTCG AACCGGCGGT ACCGGGTTGA ACAAATAACG TCGAATATTA CCAATGTTTA TTTCGTTATC GTAGTGTTTA

rmaI
bsmI maeI
3201 TTCACAAATA AAGCATTTT TTCACTGCAT TCTAGTTGTG GTTGTGCCAA ACTCATCAAT GTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC
AAGTGTTTAT TTCGTAAAAA AAGTGACGTA AGATCAACAC CAACACAGGT TGAGTAGTTA CATAGATAG TACAGACCTA GCTAGCCCTT AATTAGCCG

rmaI
haeI
styI
ncol
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
nlaIV maeI hincII/hindII
bstYI/xhoII accI pStI
bamHI xbaI pleI bspMI
alwI(dam-) hinfI bspMI
3301 GCAGGACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTAGGTAC CTTCTGAGGC GGAAGAACC AGCTGTGGAA TGTGTGTGAG TTAGGTGTG
CGTCGTGGTA CCGGACTTTA TTGGAGACTT TCTCCTTGAA CCAATCCATG GAAGACTCCG CCTTCTTGG TCGACACCTT ACACACAGTC AATCCACAC

```

SUBSTITUTE SHEET (RULE 26)

FIG. 61

[illegible]

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FIG. 6H

```

          eam1105I
          sau96I
          scrFI
          mvaI      avaII
          ecoRII
          dsav
          bstNI    asuI      mboII mboII
          bsaJI    mnlI      bpuAI earI/ksp632I   styI
          apyI(dcm+) bbsI mnlI      bsaJI
          2401 CTGTGTGACAC ACCTCCCCCA TGCCACACCGT GCCCAGCACC TGAACCTCCTG GGAGGACCGT CAGTCTTCCT CTTCCTCCCA AAACCCAAGG ATACCCCTTAT
              GAACACTGTG TGGAGGGGGT ACGGGTGCCA CGGGTGTGG ACTTGAGGAC CCTCCTGGCA GTCAGAAGGA GAAGGGGGGT TTGCGGTTCC TATGGGAATA

          sau96I
          nlaIV
          avaII
          asuI      maeII
          mspI      pmlI
          hpaII     eco72I
          scrFI     mnlI bsaAI
          nciI      ddel maeIII
          dsav      eco81I bbrPI
          cauII     bsu36I/mstII/sauI      maeII
          2501 GATTTCCCGG ACCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG AAGACCCCGA GGTCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT
              CTAAAGGGCC TGGGGACTCC AGTGCACGCA CCACCACCTG CACTCGGTGC TTCTGGGGCT CCAGGTCAAG TTCACCATGC ACCTGCCGCA CCTCCACGTA

          acil
          thal
          fnuDII/mvnI
          bstUI
          bsh1236I
          sacII/sstII
          nspBII
          kspI
          dsal
          bsaJI
          acil
          fnu4HI mnlI      maeII
          2601 AATGCCAAGA CAAAGCCCGG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT GGTCCGTC CTCACCGTC CTACCGTCG TGCACCAAGGA CTGGCTGAAC GGCAAGGAGT
              TTACGGTTCT GTTTCGGGCG CCTCCTCGTC AAGTTGTCGT GCAAGGCACA CCAGTCCGAG GAGTGGCAGG ACGTGGTCTT GACCGACTTG CCGTTCTCTCA

          scrFI
          mvaI bsrI
          ecoRII
          dsav
          bstNI
          mnlI      ecoNI bstNI
          hgaI hphI bslI apyI(dcm+)
          rsaI
          csp6I
          rsaI
          csp6I

```

FIG. 6G

hgiAI/aspHI bsp1286 bsiHRAI bmyI apaLI/snoI alw44I/snoI
 hgiAI/aspHI scrFI mvaI ecorII dsav bstNI apyI[dcM+] draIII
 bsp1286 bsiHRAI mvaI ecorII dsav bstNI apyI[dcM+] draIII
 sau96I auaII bmyI mnlI asuI apaLI/snoI bstNI apyI[dcM+] draIII
 mnlI alwNI fnu4HI mnlI nlaIV alw44I/snoI apyI[dcM+] draIII
 bsri muni bbyI mnlI nlaIV alw44I/snoI apyI[dcM+] draIII
 2001 CGGCATTATT GGAGTGAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG CCTCAATGGG ACGGTGCACC TCTCTGCCA GGAGAAACAG AACACCGTGT
 GCCGTAATAA CTCACCTTTT GGAAAAGGTC ACGAAGTTAA CGTCGGAGAC GGAGTTACCC TGGCAGGTGG AGAGGACGGT CCTCTTTGTC TTGTGGCACA
 hgiAI/aspHI bsp1286 bsiHRAI bmyI
 gsul/bpmI scrFI mvaI apaLI/snoI ecorII dsav
 bstNI alw44I/snoI apyI[dcM+] draIII
 2101 GCACCTGCCA TGCAGGTTTC TTCTTAAGAG AAAACGAGTG TGCTCTCTGT AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCACAGAT
 CGTGGACGGT ACGTCCAAAG AAAGATTCTC TTTTGCTCAC ACAGAGGACA TCATTGACAT TCTTTTCGGA CTCACGTGC TTCAACACGG ATGGGGTCTA
 aluI sstI sacI hgiJII hgiAI/aspHI eci136II bsp1286 bsiHRAI bmyI banII
 maeIII hphi draIII
 nlaIV pleI hgiCI mnlI ddeI hinfI banI
 2201 TGAGAAATGTT AAGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG AGCTCAAAAC CCCACTTGGT GACACAATC ACACATGCC ACCGTGCCA
 ACTCTTACAA TTCCCGTGAC TCCTGAGTCC GTGGTGCTG TTCTCTCAAC TCGAGTTTG GGGTGAACCA CTGTGTTGAG TGTGTACGG TGCCACGGGT
 bsp1286 nlaIV hgiJII hgiCI bsp1286 bmyI bsiHRAI bmyI banII
 maeIII mnlI nlaIV hgiJII bsp1286 bmyI bsiHRAI bmyI banII
 2301 GAGCCCAAT CTGTGACAC ACCTCCCCCG TGCCACGGT GCCCAGAGCC CAAATCTTGT GACACACCTC CCCCATGCC ACCGTGCCA GAGCCCAAT
 CTCGGGTTA GAACACTGTG TGGAGGGGGC ACGGTGCCA CGGGTCTCG GTTTAGACA CTGTGTGGAG GGGGTACGG TGCCACGGGT CTCGGGTTA

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FIG. 6F

```

scrFI
nciI
mspI
hpaII
dsav
cauII
xmaI/pspAI
smaI
scrFI
nciI
dsav
cauII
bslI
sau96I
haeIII/palI
asuI
scrFI
mvaI bsaJI
ecorII
dsav
bsrNI bsaJI
bslI avaI
apyI(dcm+)
bspl407I
rsal
csp6I
bspl407I
1801 CCTACTTGTA CAATGACTGT
GGATGAACAT GTTACTGACA GGTCCGGGCC CCGTCTATG CCTGACGTCC CTCACACTCT CGCCGAGGAA GTGGCGAAGT CTTTGGTGG AGTCTGTGAC

nlaIV
fnu4HI
aciI
acII
hphI eco57I
bsrBI
1801 CCTACTTGTA CAATGACTGT
GGATGAACAT GTTACTGACA GGTCCGGGCC CCGTCTATG CCTGACGTCC CTCACACTCT CGCCGAGGAA GTGGCGAAGT CTTTGGTGG AGTCTGTGAC

alwNI
ddeI
mnlI

scrFI
mboII
earI/ksp632I
sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
bstYI/xhoII
bglII
1901 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT CTTCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCATGAC
GGAGTCGACG AGGTTTACGG CTTTCCTTTA CCAGTCCAC CTCTAGAGAA GAACGTGTCA CTTGGCCCTG TGCCACACAC CGACGTCTT CTTGGTCATG

fnu4HI
aluI
pvuII
nspBII
ddeI
mnlI bbvI
scrFI
nciI
mspI
hpaII
dsav
cauII
sau96I
avaI
asuI
draIII
bbvI mboII bsrI cfr10I
mspI
hpaII
rsal
csp6I
fnu4HI
bsgI
psti
scfI
1901 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT CTTCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCATGAC
GGAGTCGACG AGGTTTACGG CTTTCCTTTA CCAGTCCAC CTCTAGAGAA GAACGTGTCA CTTGGCCCTG TGCCACACAC CGACGTCTT CTTGGTCATG

```

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FIG. 6E

FIG. 6E

haeIII/palI
eaeI
cfrI
mspI
hpaII
scrFI
nciI
ecoRI dsaV
taqI apoI cauII
claiI/bsp106 bsaJI aluI

scfI foki
11501 CACTATAGAA TAACATCCAC TTTGCTTTTC TCTCCACAGG TGTCACATCCA GGTCACACTGC ACCTCGGTTT TATCGATTGA ATTCCCGGC CATAGCTGTC
GTGATATCTT ATTGTAGGTG AAACGGAAAG AGAGGTGTCC ACAGTGAGGT CCAGTTGACG TGGAGCCAAAG ATAGCTAACT TAAGGGGCGG GTATCGACAG

gsul/bpmI
scrFI
mvaI
ecoRII
dsaV
bstNI
apyI[dcM+]
gsul/bpmI
maeIII hincII/hindII
bsaJI
11601 TGGCATGGGC CTCTCCACCG TGCCGTGACCT GCTGCTGCCG CTGCTGTCCC TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT
ACCGTACCCG GAGAGGTGGC ACGGACTGGA ACGGACGGC GACCACGAGG ACCTCGACAA CCACCTTAT ATGGGGAGTC CCCAATAACC TGACCAGGGA

nmII
haeIII/palI
sau96I
asuI
nlaIII
11701 CACCTAGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAATA TATCCACCTT CAAAATAATT CGATTGCTG TACCAAGTGC CACAAGGAA
GTGGATCCCC TGTCCCTCTT CTCTCTATCA CACACAGGG TTCCTTTTAT ATAGTGGGA GTTTTATTAA GCTAAACGAC ATGGTTCAG GTGTTTCCTT

nmII
ecoNI
sau96I
nlaIV
avaII
bsu36I/mstII/sauI
bslI
bsrI bslI
11801 TGGCATGGGC CTCTCCACCG TGCCGTGACCT GCTGCTGCCG CTGCTGTCCC TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT
ACCGTACCCG GAGAGGTGGC ACGGACTGGA ACGGACGGC GACCACGAGG ACCTCGACAA CCACCTTAT ATGGGGAGTC CCCAATAACC TGACCAGGGA

rmal
mael
styI
bsaJI
blnI
avrII
11901 CACCTAGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAATA TATCCACCTT CAAAATAATT CGATTGCTG TACCAAGTGC CACAAGGAA
GTGGATCCCC TGTCCCTCTT CTCTCTATCA CACACAGGG TTCCTTTTAT ATAGTGGGA GTTTTATTAA GCTAAACGAC ATGGTTCAG GTGTTTCCTT

styl
mboII
earI/ksp632I
bsaJI
12001 TGGCATGGGC CTCTCCACCG TGCCGTGACCT GCTGCTGCCG CTGCTGTCCC TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT
ACCGTACCCG GAGAGGTGGC ACGGACTGGA ACGGACGGC GACCACGAGG ACCTCGACAA CCACCTTAT ATGGGGAGTC CCCAATAACC TGACCAGGGA

rsal
csp6I
12101 CACCTAGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAATA TATCCACCTT CAAAATAATT CGATTGCTG TACCAAGTGC CACAAGGAA
GTGGATCCCC TGTCCCTCTT CTCTCTATCA CACACAGGG TTCCTTTTAT ATAGTGGGA GTTTTATTAA GCTAAACGAC ATGGTTCAG GTGTTTCCTT

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FIG. 6D

```

      haeIII/palI
      haeI
      nlaIII
      scrFI      mvaI      mboI/ndeII(dam-)      sau3AI
      mvaI      ecorII      dsav      pleI      dpnI(dam+)
      bstNI      nlaIII      bstNI      ddeI      dpnII(dam-)
      apyI(dcm+)      hinfI      apyI(dcm+)      hinfI      maeIII      alwI(dam-)
      TGGTATAGTCG      GAGGCAGTTC      TGTATTACCAG      GAAGCCATGA      ATCAACCAGG      CCACCTTAGA      CTCTTTGTGA      CAAGGATCAT
      GTTCATTTC      TCTGTACCA      ACCTATCAGC      CTCCGTCAG      ACAATGGTC      CTTCGGTACT      TAGTTGGTCC      GGTGGATCT      GAGAACACT      GTTCCTAGTA
      accI      nlaIII      mnlI
      1101 CAAGTAAAGT      AGACATGGTT      TGGATAGTCG      GAGGCAGTTC      TGTATTACCAG      GAAGCCATGA      ATCAACCAGG      CCACCTTAGA      CTCTTTGTGA      CAAGGATCAT
      GTTCATTTC      TCTGTACCA      ACCTATCAGC      CTCCGTCAG      ACAATGGTC      CTTCGGTACT      TAGTTGGTCC      GGTGGATCT      GAGAACACT      GTTCCTAGTA

      maeII
      aflIII
      apoI      maeIII
      1201 GCAGGAATTT      GAAAGTGACA      CGTTTTTCCC      AGAAATTGAT      TTGGGGAAT      ATAAACCTCT      CCCAGAATAC      CCAGGCGTCC      TCTCTGAGGT      CCAGGAGGAA
      CGTCCCTTAA      CTTTCACTGT      GCAAAAAGGG      TCTTTAACTA      AACCCTTTTA      TATTGGAGA      GGGTCTTATG      GGTCCGAGG      AGAGACTCCA      GGTCTCTCTT

      mnlI
      hinfI/acyI      scrFI
      ahaII/bsaHI      mvaI
      scrFI      ecorII      dsav
      mvaI      ecoNI      sau96I
      dsav      avaII
      bstNI      bslI      asuI      mnlI
      apyI(dcm+)      mnlI      bstNI
      bsaJI      hgaI      ddeI      apyI(dcm+)
      1201 GCAGGAATTT      GAAAGTGACA      CGTTTTTCCC      AGAAATTGAT      TTGGGGAAT      ATAAACCTCT      CCCAGAATAC      CCAGGCGTCC      TCTCTGAGGT      CCAGGAGGAA
      CGTCCCTTAA      CTTTCACTGT      GCAAAAAGGG      TCTTTAACTA      AACCCTTTTA      TATTGGAGA      GGGTCTTATG      GGTCCGAGG      AGAGACTCCA      GGTCTCTCTT

      sfaNI      mboII      mnlI      aluI
      1301 AAAGGCATCA      AGPATAAGTT      TGAAGTCTAC      GAGAAGAAAG      ACTAACAGGA      AGATGCTTTC      AAGTTCTCTG      CTCCCTCTCT      AAAGCTATGC      ATTTTATATA
      TTCCGCTAGT      TCATATTCAA      ACTTCAGATG      CTCTTCTTTC      TGATTGCTCT      TCTACGAAAG      TTCAAGAGAC      GAGGGAGGA      TTTCGATACG      TAAAAATATT

      fnu4HI
      aciI
      thaI
      fnuDII/mvnI      tru9I
      bstUI      mseI
      bsh1236I      aseI/asnI/vspI
      styI      bsaJI
      1401 GACCATGGGA      CTTTGTCTGG      CTTTAGACCC      CTTTGGCTTC      GTTAGAACGC      GGCTACAATT      AATACATAAC      CTTATGTATC      ATACACATAG      ATTTAGGTGA
      CTGGTACCCT      GAAAACGACC      GAAATCTGGG      GGAACCGAAG      CAATCTTGGG      CCGATGTTAA      TTATGTATTG      GAATACATAG      TATGTATC      TAAATCCACT

```


FIG. 6C

tfil
 acil
 thaI hinfI
 fnuDII/mvnI
 bstUI
 bsh1236I
 701 TTGGAACGG GATTCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA GCGATAAGAG GATTATATCC CGCTGCCAT CATGTTTGA CCATTGAAC
 AACCTTGGC CTAAGGGCA CGTTCTCAC GACATTCTAC GCGGATATCT CGCTATTCTC CTAAATAGG GCGACAGCT GTACCAAGCT GGTAACTTGA
 fnu4HI
 bbvI
 nspBII
 acil
 nlaIII
 taqI
 thaI
 fnuDII/mvnI
 bstUI
 bsh1236I
 mlui
 bsrBI
 aflIII
 rsaI
 acil
 xmnI
 csp6I
 mnli
 ddel
 asp700
 scaI
 801 GCATCGTGC CGTGTCCTCA AATATGGGA TTGGCAAGAA CGGAGACCTA CCCTGCCCTC CGCTCAGGAA CGCTTCAAG TACTTCCAA GAATGACCAC
 CGTAGCAGG GCACAGGTT TTATACCCCT AACCGTTCTT GCCTCTGGAT GCGACGGGAG GCGAGTCTT GCGCAAGTTC ATGAAGTTT CTTACTGGTG
 pflMI
 bslI
 sfaNI
 eco57I
 mboII
 earI/ksp632I
 mnli
 901 AACCTCTTCA GTGGAAGGTA AACAGATCT GGTGATTATG GGTAGGAAA CCTGGTTCTC CATTCCTGAG AAGATCGAC CTTTAAAGGA CAGAAATTAAT
 TTGGAGAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAAATTTCT GTCTTAATTA
 tfil
 hinfI
 alwNI
 hphI
 mboII
 taqI
 msei
 trui
 msei
 ahaIII/draI
 asei/asnI/vspI
 aluI
 sstI
 sacI
 hgiJII
 hgiAI/aspHI
 ecl136II
 bsp1286
 bsiHKAI
 bmyI
 banII
 1001 ATAGTTCTCA GTAGAGAACT CAAGAACCA CCACGAGGAG CTCATTTCTT TGCCAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTCG
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCTC GAGTAAAGA ACGGTTTCA AACCTACTAC GGAATCTCA ATAATTTGT GGCCTTAACC
 ddel
 bslI
 mnli
 bstXI
 foki
 sfanI
 msei
 afliI/bfri
 bsaNI
 mspI
 hpall

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FIG. 6B

401 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGA TTTCAGTCAT TGACGCCAT GGGAGTTGT TTTGGCACCA
 CCAAAACCGT CATGTAGTTA CCGCACCTA TCGCCAACT GAGTGGCCCT AAAGTTTCAG AGGTGGGTA ACTGCAGTTA CCTCAAACA AAACCGTGTG

rsal csp6I pleI
 maeII hinII/acyI nlaIV
 ahaII/bsaHI hgiCI
 aatII bani

501 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC AAATGGCGG TAGGCGTGTG CGTGGGAGG TCTATATAAG CAGAGCTCGT
 TTTAGTTGCC CTGAAAGGTT TTACAGCATT GTTGAGGCGG GGTAACCTGG TTTACCCGCC ATCCGCACAT GCCACCCCTCC AGATATATTC GTCTCGAGCA

maeIII aciI hgaI
 rsal csp6I mnlI
 aciI csp6I
 hgiJII
 hgiAI/aspHI
 eclI36II
 bspI286
 bsiHRAI
 bmyI
 banII

601 TTAGTGAACC GTCAGATGCG CTGGAGACGC CATCCACGCT GTTTTGACCT CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA
 AATCACTTGG CAGCTTAGCG GACCTCTGCG GTAGGTGCGA CAAACTGCGA GGTATCTTCT GTGGCCCTGG CTAGGTGGA GCGCGCCGCC CTTCACACGT

esp3I
 scrFI
 mvaI bsmAI
 ecorII
 dsav
 bstNI hinII/acyI
 apyI(dcm+)
 sau3AI gsuI/bpmI
 mboI/ndeII(dam-)
 dpnI(dam+) hgaI fokI
 dpnII(dam-) ahaII/bsaHI
 mnlI
 cauII
 bbsI
 mboII hpaII
 bpuaI dsav
 mspI
 nciI
 scrFI
 nlaIV
 asuI
 sau96I
 avall
 kspI scrFI
 dsal nciI
 bglI bslI mspI
 sau3AI mnlI bstUI
 mboI/ndeII(dam-) hpaII
 dpnI(dam+) bsaJI dsav
 dpnII(dam-) bshI236I
 alwI(dam-) aciI cauII
 GATCCAGCCT CCGCGGCCCG GAACGGTGCA
 CTAGGTGGA GCGCGCCGCC CTTCACACGT

haeIII/palI
 mcrI
 eagI/xmaIII/eclXI
 eaeI
 cfrI
 fnu4HI
 aciI
 thaI
 fnuDII/mvnI
 sacII/sstII
 nspBII
 kspI scrFI
 dsal nciI
 bglI bslI mspI
 sau3AI mnlI bstUI
 mboI/ndeII(dam-) hpaII
 dpnI(dam+) bsaJI dsav
 dpnII(dam-) bshI236I
 alwI(dam-) aciI cauII
 GATCCAGCCT CCGCGGCCCG GAACGGTGCA
 CTAGGTGGA GCGCGCCGCC CTTCACACGT

FIG. 6A

[illegible]

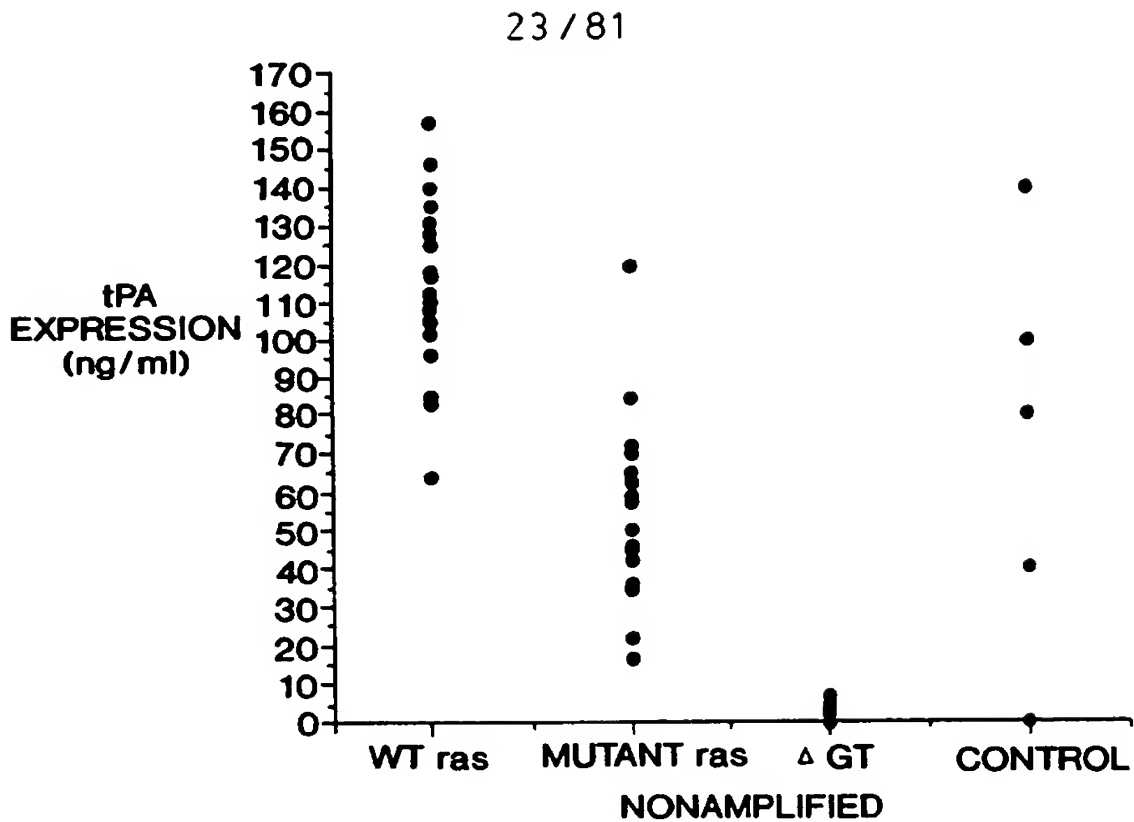
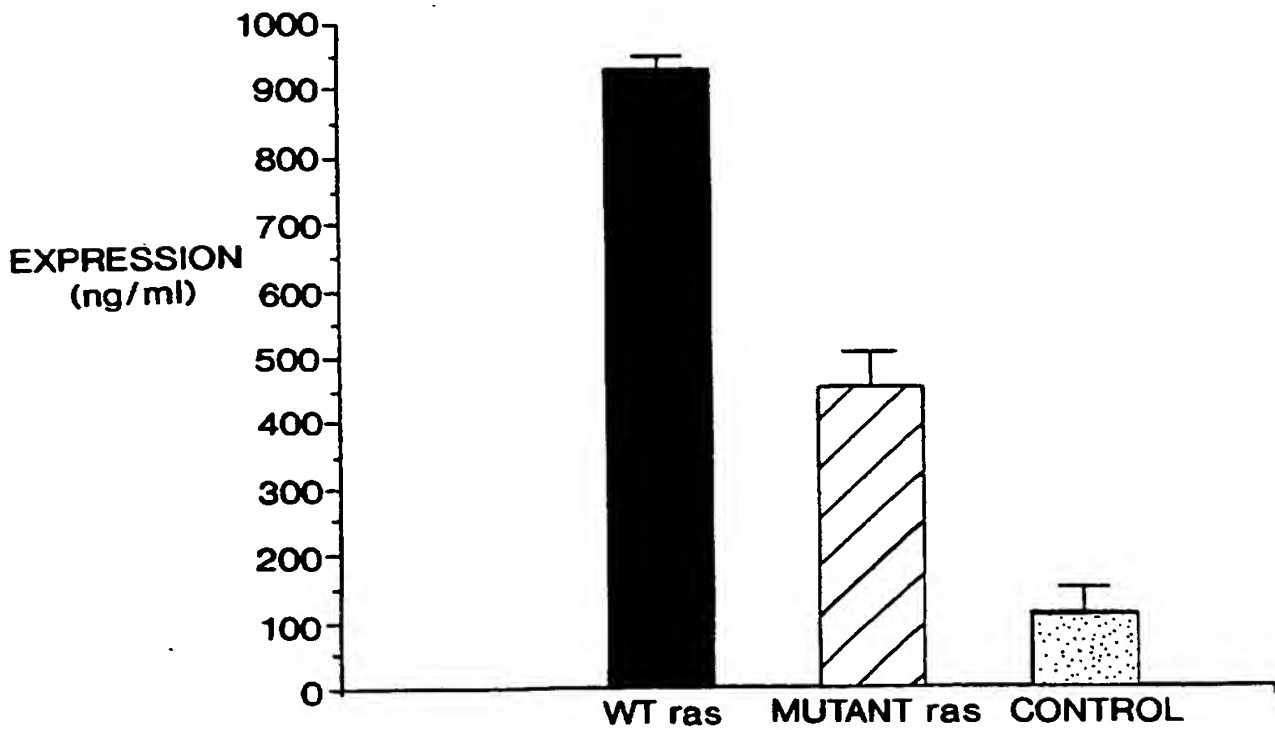


FIG. 5B



AMPLIFIED (200nM METHOTREXATE)

FIG. 5C

FIG. 10B

401 GCTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA TTTCACAGTC TCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA
 CCAAAACCGT CATGTAGTTA CCCGCACCTA TCGCCAAACT GAGTGCCCT AAAGTTTCA AGGTGGGTA ACTGCAGTTA CCTCAAACA AAACCGTGT
 rsal maeIII hinII/acyI pleI aciI hinfi bsmAI
 csp6I
 501 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC AATGGGCGG TAGCGTGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT
 TTTAGTTGCC CTGAAAGGTT TTACAGCAAT GTTGAGGCGG GGTAACCTCGG TTTACCCGCC ATCCGCACAT GCCACCCTCC AGATATATTC GTCTCGAGCA
 maeIII aciI hgaI hgaIII/pali
 csp6I
 601 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT CCATAGAAGA CACCGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA
 AATCACTGG CAGTCTAGCG GACCTCTGCG GTAGTGCGA CAAAACTGGA GGTATCTTCT GTGGCCCTGG CTAGTTCGA GCGCGCGGCC CTTGCCACGT
 esp3I
 scrFI
 mvaI bsmAI
 ecorII
 dsav
 bstNI hinII/acyI
 apyI[dcn+]
 sau3AI gsuI/bpmI
 mboI/ndeII[dam-]
 dpnII[dam+] hgaI fokI
 dpnII[dam-] ahaII/bsaHI
 sau96I
 asuI
 nlaIV
 scrFI
 nciI
 mspI
 hpaII
 dsav
 mboII
 bpuAI
 bbsI
 mnlI
 cauII
 sacII/sstII
 nspBII
 kspI scrFI
 dsal nciI
 bglI bslI mspI
 sau3AI mnlI bstUI
 mboI/ndeII[dam-] hpaII
 dpnII[dam+] bsaJI dsav
 dpnII[dam-] bsh1236I
 alwI[dam-] aciI cauII
 fnuDII/mvni
 fnu4HI
 aciI
 thaI
 mcrI
 eagI/xmaIII/ecI XI
 eaeI
 cfrI
 bmyI
 bsiHKA
 bsp1286
 ecII36II
 hgiAI/aspHI
 hgiJII
 sacI
 sstI
 aluI
 bani
 hgiCI
 nlaIV

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Fig. 100

```
tfII      fnu4HI
aciI      aciI
thAI      thAI
fnuDII/mvniI  bstUI
bstUI      bstUI
bsh1236I  bsh1236I
701 TTGGAACGCG GATTCCCGGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA GTCTATAGGC CCACCCCTT GGCTCGTTA GAACGGGCTT ACAAATTAATA
AACCTTGGCG CTAGGGGCA CGGTTCTCAC TGCATTTCATG GCGGATATCT CAGATATCCG GGTGGGGGAA CCGAAGCAAT CTTCGCCCGA TGTTAATTAT
      sau96I
      avaiI
      asuI
      scrFI
      mvaI
      ecoRII
      dsav
      bstNI
      apyI{dcm+}
      bslI bsaJI
801 CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA CATCCACTTT GCCTTTCTCT CCACAGGTGT CCATCCCCAG GTCCAACCTGC
GTATTGGAAT ACATAGTATG TGTATGCTAA ATCCACTGTG ATATCTTATT GTAGGTGAAA CGGAAGAGA GGTGTCCACA GGTGAGGTC CAGGTTGACG
      hinII/acyI
      ahaII/bsaHI
      aatII
      thAI
      fnuDII/mvniI
      bstUI
      aciI maeII
      hphi bsh1236I taqI
      aluI taqI
      hindIII
      claI/bspl06
      bsaJI ddeI
901 ACCTCGGTTT TAAGCTTATC GATATGAAAA AGCCTGAACT CACCGCGACG TCTGTGAGA AGTTTCTGAT CGAAAGTTC GACAGGCTCT CCGACCTGAT
TGGAGCCCAAG ATTGGAATAG CTATACTTTT TCGGACTTGA GTGGCGCTGC AGACAGCTCT TCAAAGACTA GCTTTTCAAG CTGTCCGAGA GGCTGGACTA
      hinPI
      fnu4HI
      bbvI
      aluI
      tfII
      hinfi
      mnlI mboII
      aluI
      taqI
      aluI
      mnlI
      aciI
      hhaI/cfoI
      bbvI
1001 GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTAGCTTC GATGTAGGAG GGCGTGGATA TGTCTCTCGG GTAATAGCT CGCGCGATGG TTTCTACAAA
CGTCGAGAGC CTCCCGCTTC TTAGAGCAGG AAGTCTGAAG CTACATCTTC CCGCACCTAT ACAGGACGCC CATTTATCGA CGCGGCTACC AAAGATGTTT
```

FIG. 10D

hinPI mspI hhaI/cfoI hpaI
 thai acil mroI
 haeIII/palI bspMI
 mcrI fnuDII/mvni bspEI
 eagi/xmaIII/ecI XI bsaWI
 eaeI bstUI tfil
 cfrI bsh1236I hinfi
 sfaNI fnu4HI bslI accIII
 mboI/ndeII(dam-) ecorI
 dpnI(dam+) apoI
 dpnII(dam-) sfaNI acil
 1101 GATCCTTATG TTTATCGGCA CTTTGCATCG GCGCGCTCC CGATTCCGGA AGTGCTTGAC ATTGGGAAT TCAGCGAGAG CTGACCTAT TGCATCTCCC
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGCGCGGAGG GCTAAGGCCT TCACGAACTG TAACCCCTTA AGTCGCTCTC GGACTGGATA ACGTAGAGGG

mcrI eagi/xmaIII/ecI XI
 eaeI fnu4HI
 styI
 thaI ncoI sau3AI acil
 fnuDII/mvni mboI/ndeII(dam-)
 fnu4HI bstUI dsal dpnI(dam+) sau3AI
 bbvI mcrI haeIII/palI dpnII(dam-) mboI/ndeII(dam-)
 scfI mspI bsh1236I bsaJI sfaNI fnu4HI dpnI(dam+)
 pstI hpaII mnli nlaIII pvuI/bspCI haeIII/palI
 bsgI cfr10I acil haeI foki mcrI bbvI cfrI dpnII(dam-)
 nspBI acil
 1201 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAC CGAACTGCCC GCTGTTCTGC AGCGGTGCG GGAGGCCATG GATCGGATCG CTGCGGCGGA,
 CGGCACGTCT CCCACAGTGC AACGTTGTGG ACGGACTTGG GCTTGACGGG CGACAAGAGC GTCGCGTAC CTACGCTAGC GACGCCGCT

sau96I
 avaiI
 asuI
 sau96I xsrII/cspI
 haeIII/palI acil tfil
 asuI cpoI hinfi
 acil bsrBI
 ddeI
 1301 TCTTAGCCAG ACGAGCGGT TCGGCCCAT TCGAGCCGCA GGAATCGGTC AATACACTAC ATGGCGTAT TTCATATCGG CGATTGCTGA TCCCATGTG
 AGAATCGGTC TGCTCGCCA AGCCGGGTAA GCCTGGCGTT CCTTAGCCAG TTATGTGATG TACCGCACTA AAGTATACGC GCTAACGACT AGGGGTACAC

hinPI hhaI/cfoI
 thaI
 fnuDII/mvni
 bstUI bsh1236I taqI aluI
 bsh1236I hgaI
 drdI
 1401 TATCACTGGC AAACGTGTGAT GGACGACACC GTCACTGCGT CCGTCGCGCA GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGACTGCCCC GAAGTCCGCG
 ATAGTGACCG TTTGACACTA CCTGCTGTGG CAGTCACGCA GGCAGCGCGT CCGAGAGCTA CTCGACTAGC AAACCCGCT CCGACGCGG CTTCAGGCGG

draIII
 nlaIV
 hgiCI
 bsaJI
 haeIII/palI
 sau96I
 sfaNI
 asuI
 bslI
 hpaII
 mspI

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FIG. 10E

aciI
 thal
 fnuDII/mvnI
 hgiAI/aspHI
 bsp1286
 bsiHKA1
 bmyI bstUI
 apaLI/snoI
 alw44I/snoI
 mnlI bsh1236I nlaIV
 1501 ACCTCGTGCA CGCGGATTTC GGCTCCAACA ATGTCTCTGAC GGACAATGGC CGCATACAG CCGTCATTGA CTGGAGCGAG GCGATGTTTC GGGATTCCCA
 TGGAGCACGT GCGCCTAAG CCGAGGTTGT TACAGGACTG CCGTTTACCG GCGTATTGTC GCGAGTAACT GACCTCGCTC CGCTACAAGC CCCTAAGGGT
 aciI
 fnu4HI
 haeIII/palI
 eaeI
 cfrI
 nspBII
 aciI
 gsuI/bpmI
 bsrI
 mnlI
 tfil bslI
 hinfi
 fnu4HI
 thal
 fnuDII/mvnI
 bstUI
 bsh1236I
 sacII/sstII
 nspBII
 kspI
 dsal
 bsaJI
 aciI
 fnu4HI
 sau3AI aciI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 alwI(dam-)
 mspI
 hpaII
 mroI
 bspMII
 bspEI
 bsaWI
 rsaI
 aciI
 foki
 csp6I
 bsrBI
 sfanI
 aluI
 fnu4HI
 bbvI
 maeII
 taqI
 mnlI
 accIII
 1601 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTCTATGG AGCAGCAGAC GTACTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG
 TATGCTCCAG CGGTTGTAGA AGAAGACCTC CGSCACCAAC CGAACATACC TGTGCTCTG CATGAAGCTC GCCTCCGTAG GCCTCGAAGC TCCTAGCGGC
 dsal
 haeIII/palI
 mboII mnlI bsaJI
 mboII gsuI/bpmI
 mnlI
 1701 CGGCTCGGG CGTATATGCT CCGCATTTGGT CTTGACCAAC TCTATCAGAC CTTGGTTGAC GGCAATTTTC ATGATGCAGC TTGGCGGCAG GGTGATCGG
 CGCCAGGCC GCATATACGA GCGGTAACCA GAAGTGGTTG AGATAGTCTC GACCAACTG GACCAACTG TACTAGCTC AACCGCGCTC CCAGCTAGC
 nlaIV
 scrFI
 nciI
 mspI
 hpaII
 dsav
 cauII
 aluI
 nincII/hindII
 taqI
 sfanI
 bbvI
 fnu4HI
 aluI
 hinPI
 taqI
 drdI
 hgaI
 hhaI/cfoI
 sfanI

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FIG. 10F

```

nlaIV
mspI      haeIII/palI
hpaII     mcrI
bslI      eagI/xmaIII/ecI XI
mroI      eaeI
bspMII    cfrI
bspEI(dam-) fnu4HI
bsaWI     aciI
accIII(dam-) thal
sau3AI    fnuDII/mvmI
mboI/ndeII(dam-) bstUI
dpmI(dam+) bsh1236I
dpmII(dam-) hinPI
alwI(dam-) rsal
                  csp6I
1801 ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAATCGCC CGCAGAAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAG TACTGCCGA
TCCGTTAGCA GGCTAGGCCT CGGCCCTGAC AGCCCGCATG TGTTAGCGG CGGTCTTGC GCCGCGAGAC CTGGCTACCG ACACATCTTC ATGAGCGGCT
                  aciI
                  hhaI/cfoI
                  asuI
                  scaI
                  rsal
                  csp6I
                  scaI
scrFI
ncII
mspI
hpaII
dsav
xmaI/pspAI
smaI
scrFI
ncII
dsav
cauII
bsaJI
aval
bsaJI
sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
nlaIV cauII
bstVI/xhoII
bamHI bsaJI ecoRI
alwI(dam-) apoI
cIal/bsp106 bsaJI
mcrI
bslI
sfaNI
mnII
bsaJI
hinII/acyI
hgaI
ahaII/bsaHI
1901 TAGTGGAAAC CGACGCCCCA GCACCTCGTCC GAGGGCAAAG GAATAGAGTA GATGCCGACC GAAGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC
ATCACCTTTG GCTCGGGGT CGTGAGCAGG CTCCCGTTTC CTTATCTCAT CTACGGCTGG CCTTCTAGGG GCCCCTTAAG TTAGCTACCG GCGGTACCGG
                  aciI
                  fnu4HI
                  bglI nlaIII
                  sfiI styI
                  eaeI ncoI
                  cfrI dsal
                  taqI haeIII/palI
                  cIal/bsp106 bsaJI
                  AATCGATGGC CGCCATGGCC
                  TTAGCTACCG GCGGTACCGG
```

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FIG. 10G

2001 CAACCTGTTT ATTCAGCTT ATAATGGTTA CAATAAAGC AATAGCATCA CAATTCAC AATAAAGCA TTTTTCAC TGCATTCAG TTGCGGTTG
 GTTGACAAA TAACGTCGAA TATTACCAAT GTTATTTCG TTATCGTAGT GTTTAAGTG TTTATTTCGT AAAAAAAGTG ACGTAAGATC AACACCAAC
 aluI fnu4HI bbvI maeIII sfanI apoI rnaI bsmI maeI
 sau3AI mboI/ndeII(dam-) dpnI(dam+) dpnII(dam-) pvuI/bspCI mcrI
 taqI(dam-) tru9I claiI/bsp106(dam-) sau3AI mseI fnu4HI styI haeIII/palI haeI
 mboI/ndeII(dam-) dpnI(dam+) xmnI hinPI dsal bbvI ncoI
 dpnII(dam-) asel/asnI/vspI bsaJI
 nlaIII alwI(dam-) asp700 hhaI/cfoI nlaIII mnlI
 2101 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGGAAATTAAT TCGGGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGTTA
 AGGTTTGAGT AGTTACATAG AATAGTACAG ACCTAGCTAG CCCTTAATTA AGCCGCTCG TGGTACCAG CTTTATTGGA GACTTCTCCTC TTGAACCAAT
 rsaI csp6I nlaIV kpnI hgiCI banI aluI pvuII nspBII
 asp718 mnlI acc65I ddeI aciI
 2201 GGTACCTTCT GAGCGGAAA GAACCAAGCTG TGGAAATGCT GTGCGAAG GTGCGAAG TCCCCAGGT TCCCCAGGT CAGAAAGTATG CAAAGCATGC
 CCATGGAAGA CTCGCGCTTT CTTGGTCGAC ACCTTACACA CAGTCAATCC CACACCTTTC AGGGGTCCGA GGGGTCTGCC GTCTTCATAC GTTTCGTACG
 nlaIV
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI
 2301 ATCTCAATTA GTCAGCAACC AGGTGTGGA AGTCCCCAGG CTCCCCAGCA GGCAGAAGTA TGCAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC
 TAGAGTTAAT CAGTCGTTGG TCCACACCTT TCAGGGGTCC GAGGGGTCTC CCGTCTTCAT ACCTTTCGTA CGTAGAGTTA ATCAGTCGTT GGTATCAGGG
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI sexAI
 sphI nspi sfanI nspHI aciI

FIG. 10H

[illegible][illegible][illegible]

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FIG. 10I

```

sau3AI      sau96I
mboI/ndeII(dam-)  avaiI
dpnI(dam+)      asuI
dpnII(dam-)     scrFI
alwI(dam-)      mvaI
taqI(dam-)      ecoRII
claiI/bsp106(dam-)  dsav
sau3AI          bstNI
mboI/ndeII(dam-)  apyI(dcm+)
dpnI(dam+)      bsII bsaJI
dpnII(dam-)     foki
alwI(dam-)      bsII bsaJI
2701 ACCTTTTGGG TCGATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC CACAGGTGTC CACTCCGAGG TCCAACTGCA CCTCGGTTCC CGAAGCTAGC
TGGAAACCT AGCTAGGATG ACTGTGACTG TAGGTGAAA AGAAAAAGAG GTGTCCACAG GTGAGGGTCC AGTTGACGT GGAGCCCAAGC GCTTCGATCG

nlaIII
styI
pflMI
ncol
sfanI      ecoRI
fnu4HI taqI apoI      bsaJI foki
bbvI claiI/bsp106    nlaIII foki
2801 TTGGGGCTGCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
AACCCGACGT AGCTAACTTA AGTGGGTACC CTACCAGTAC ATAGTAGGAA AAAGATCATC GTTGACGTTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

alul
sstI
sacI
hgiJII
hgiAI/aspHI
ecII36II
bsp1286
bsiHKA1
bmyI
banII      mnlI
avaI      aciI
2901 GTCCCCGAGC TCCCTGTCCG CCTCTGTCCG CGATAGGGTC ACCATCACCT GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACCTGG
CAGGGGCTCG AGGACAGGC GGAGACACCC GCTATGCCAG TGCTAGTGGA CGGCACGGTC AGTCTCCGAG CTAATGCTAC CACTATCGAT GTACTTGACC

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FIG. 10J

mspI
 hpaII
 bslI
 bsaWI
 sau3AI
 mboI/ndeII[dam-]
 dpnI[dam+]
 dpnII[dam-]
 alwI[dam-]
 nlaIV
 bstYI/xhoII
 bamHI
 alwI[dam-]
 3001 TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACAGCGCCTC GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGGA
 ATAGTTGTCT TTGGTCTCTT TCGAGGCTTT GATGACTAAA TCGCGCGGAG CATGGACCTC AGACCTCAGG GAAGAGCGAA GAGACCTAGG CCAAGACCCCT
 gsuI/bpmI
 scrFI
 mvaI
 haeIII/palI
 fnu4HI
 ecorII
 aciI
 dsav
 thal mnlI bstNI
 fnuDII/mvnI apyI[dcn+] pleI
 bstUI
 rsaI
 pleI
 gsuI/bpmI
 bsh1236I
 csp6I
 hinFI
 hinFI
 3101 CGGATTTTCC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTCGCACT TATTACTGTC AGCAAGTCA CGAGGATCCG TACACATTG GACAGGGTAC
 GCCTAAAGTG AGACTGGTAG TCGTCAGACG TCGGCTCTCT GAAGCGTTGA ATAATGACAG TCGTTTCAGT GCTCCTAGGC ATGTGTAAC CTGTCCCATG
 fnu4HI mboII
 bbvI
 bpuAI
 scfI
 bbsI
 pstI
 mspI
 bsgI
 hpaII
 sau3AI
 mboI/ndeII[dam-]
 dpnI[dam+]
 fnu4HI
 bpuAI
 bbsI
 mboII
 3201 CAAGGTGGAG ATCAACGAA CTGTGGCTGC ACCATCTGTC TTCACTCTCC GCCATCTGA TGAGCAGTTG AAATCTGGAA CTGCTCTGT TGTGTGCTG
 GTTCCACCTC TAGTTTGCCTT GACACCGACG TGGTAGACAG AAGTAGAAGG GCGGTAGACT ACTCGTCAAC TTTAGACCTT GACGGAGACA ACACACGGAC
 haeIII/palI
 haeI
 rsaI
 mnlI
 csp6I
 xmnI
 asp700
 3301 CTGAATAACT TCTATCCAG AGAGGCCAAA GTACAGTGA AGGTGGATAA CCGCTCCAA TCGGGTAAC CCCAGAGAG TGTACAGAG CAGACAGCA
 GACTTATTGA AGATAGGTC TCTCCGGTTT CATGTACACT TCCACCTATT GCGGAGGTT AGCCCATGA GGGTCTCTC ACAGTGTCTC GTCTGTCTG

FIG. 10K

sstI
 sacI
 hgiII
 hgiAI/aspHI
 eci136II
 bsp1286
 bsiHKA1
 bmyI
 haeIII/palI
 sau96I aluI
 asuI banII
 hphI
 eco0109I/draII
 maeIII alwNI ddeI
 accI
 cgcCTGCGAA
 gTCACCCATC AGGGCCTGAG
 GCGGACGCTT CAGTGGGTAG TCCCGGACTC

ddeI
 celII/espI
 bpu1102I
 hgaI
 ddeI fnu4HI
 mnlI bbvI
 scfI
 3401 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC TACGAGAAAC ACAAGTCTA CCGCTGCGAA
 TCCTGTCTGTG GATGTGGGAG TCGTCTGTGGG ACTGGGACTC GTTCTGTCTG ATGCTCTTTG TGTTCAGAT GCGGACGCTT CAGTGGGTAG TCCCGGACTC

sau96I
 nlaIII
 aciI haeIII/palI
 fnu4HI asuI
 bglI styI
 aluI sfII ncoI
 hindIII eaeI dsal
 tru9I cfrI bsaJI
 mseI taqI haeIII/palI
 maeIII aluI
 3501 CTCGCCCGTC ACAAGAGCT TCAACAGGGG AGAGTGTAA GCTTCGATGG CCGCATGGC CCACTTGT TATTGCAGCT TATAATGTT ACAATAAAG
 GAGCGGCAG TGTCTCTCGA AGTTGTCTCC TCTCACAATT CGAAGCTACC GCGGTACCG GGTTGAACAA ATAACGTCTGA ATATTACCA TGTATTATTC

sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 pvuI/bspCI
 mcrI
 taqI(dam-)
 claI/bsp106(dam-)
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 nlaIII alwI(dam-)
 imal
 bsmI maeI
 sfanI apoI
 3601 CAATAGCATC ACAATTTC CAAATAAAGC AATTTTTTCA CTGCATTCTA GTTGTGGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT
 GTTATCGTAG TGTTTAAAGT GTTTATTTCG TAAAAAAGT GACCTAAGAT CAACACCMAA CAGGTTTGG TAGTTACATA GAATAGTACA GACCTAGCTA

FIG. 10L

[illegible]

SUBSTITUTE SHEET (RULE 26)

[illegible]

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FIG. 100

```

scrFI      thai      fnuDII/mvnI
nciI       bstUI
mspI       bsh1236I
hpaII fnu4HI hinPI
dsav aluI nspHI hhaI/cfoI
cauII bbvI nlaIII mnlI hphI hphI hphI
5001 TCCGGGAGCT GCATGTGTCA GAGGTTTCA CGTTCATCAC CGAACGCGC GAGGCAGTAT TCTTGAAGAC GAAAGGGCT CGTGATACGC CTATTTTAT
AGGCCCTCGA CGTACACAGT CTCCAAAAGT GGCAGTAGTG GCTTTGGCG CCGGTCATA AGAAGCTCTG CTTTCCCGGA GCACTATGCG GATAAAATA

nlaIV
aciI
thai
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI
maeII
hinII/acyI
ahaII/bsaHI
dclI aatII
tru9I rcaI
mseI bspHI
nlaIII
nlaII
rcaI
bspHI
bsrBI bsmAI
aciI nlaIII
5101 AGGTTAAGT CATGATAATA ATGGTTTCTT AGAGCTCAGG TGGCACTTTT CGGGGAAATG TCGCGGAAC CCCTATTGT TTATTTTCT AAATACATTC
TCCAAATTACA GTACTATTAT TACCAAGAA TCTGCAGTCC ACCGTGAAA GCCCCTTTAC AGCGCCTTG GGGATAACA AATAAAAGA TTTATGTAG

hgiAI/aspHI
bsp1286
sau3AI
mboI/ndeII(dam-)
dpmI(dam+) bmyI
dpmII(dam-)
hpaI/snoI
alw44I/snoI maeIII
5201 AAATATGTAT CCGCTCATGA GACATAACC CTGATAAATG CTCAATAAT ATTGAAAAAG GAAGAGTATG AGTATTCAC ATTTCCGTGT CGCCCTTATT
TTTATACATA GCGGAGTACT CTGTATTGG GACTATTAC GAAGTTATTA TAACTTTTC CTCTCATAC TCATAAGTTG TAAAGGCACA GCGGGAATAA

hgiAI/aspHI
bsp1286
sau3AI
mboI/ndeII(dam-)
dpmI(dam+) bmyI
dpmII(dam-)
hpaI/snoI
alw44I/snoI maeIII
5301 CCCTTTTTG CGGCAATTTG CCTTCCTGTT TTTGCTCACC CAGAAACGCT GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGTGACAGA GTGGTTACA
GGGAAAAAAC GCCGTAAAC GCGGTAAAC GGAAGGACAA AAACAGTGG GTCTTTGCGA CCACCTTTCAT TTTCTACGAC TTCTAGTCAA CCCACGTGCT CACCCAATGT

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SUBSTITUTE SHEET (RULE 25)

FIG. 10P

```
5401 sau3AI mboI/ndeII(dam-) sau3AI mboI/ndeII(mvnI)
    dpnI(,am+) dpnI(,dam-) dpnI(,dam-) bsp1286 tru9I bsh1236I
    l:stVI/xhoII nspBII alwI(dam-) alwI(dam-) bsiHKAI mseI hinPI
    bsrI alwI(dam-) aciI bstVI/xhoII mboII maeII bmyI ahaIII/draI hhaI/cfoI
    taqI TCGAACCTGGA TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA GAACGTTTC CAATGATGAG CACTTTTAA GTTCTGCTAT GTGGCGCGGT
    AGCTTGACCT AGAGTTGTG CCAATTCTAGG CAATTCTCANA AGCGGGGCTT CTTCGANAAG GTTACTACTC GTGAAAATTT CAACAGGATA CACCGCGCCA

5501 scrFI nciI mspI hpaII dsaV cauII
    hinII/acyI hgaI aciI
    ahaII/bsaHI bcgI mcrI fnu4HI ddeI
    ATTATCCCGT GATGACGCCG GGCAAGACA ACTCGTGC CGCATACACT ATTCTCAGAA TGACTTGGT GAGTACTCAC CAGTCACAGA AAGCATCTT
    TAATAGGCA CTACTGGGC CGTTCTCGT TGAGCCAGCG GGTATGTGA TAAGAGTCTT ACTGAACCAA CTCATGAGTG GTCAGTGTCT TTTCGTAGAA

5601 foki nlaIII fnu4HI bbvI nlaIII aciI
    ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG TGATAACACT GCGGCCAACT TACTTCTGAC AAGCATCGGA GGACCGAAGG
    TGCCCTACCGT ACTGTCAATC TCTTAATACG TCACGACGGT ATTGGTACTC ACTATTGTGA CGCCGGTTGA ATGAAGACTG TTGCTAGCCT CCTGGCTTCC

5701 aluI aciI nlaIII dpnII(dam-) alwI(dam-) nlaIII maeIII
    AGCTAACCGC TTTTTCGAC AACATGGGG ATCATGTAAC TCGCCTTGAT CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAACGACG AGCGTGACAC
    TCGATTGGCG AAAAAACGTG TTGTACCCCC TAGTACATTG AGCGGAACCTA GCAACCCCTG GCCTGACTT ACTTCGGTAT GGTTCCTGC TCGCACTGTG
```

FIG. 10Q

[illegible]

FIG. 10R

sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 bstYI/xhoII
 sau3AI alwI(dam-)
 mboI/ndeII(dam-)
 dpnI(dam+) mboII(dam-)
 dpnII(dam-)
 6201 GTGAAGATCC TTTTIGATAA TCTCATGACC AAAATCCCTT AACGTGAGTT TTCGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCANA GGATCTTCTT
 CACTTCTAGG AAAAATAATT AGAGTACTGG TTTTAGGGAA TTGCACTCAA AAGCAAGGTG ACTCGCAGTC TGGGGCATCT TTTCTAGTTT CCTAGAAGAA
 ddeI hgaI
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 alwI(dam-)
 mspI
 hpaII
 aluI
 6301 GAGATCCTTT TTTTCTGCG GTATCTGCT GCTTGCAAC AAAAAACCA CCGTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACCTCTTT
 CTCTAGGAA AAAAGACGCG CATTAGACGA CGAACGTTTG TTTTITGGT GCGATGCTC GCCACCAAC AAACGGCCTA GTTCTCGATG GTTGAGAAAA
 haeIII/palI
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 alwI(dam-)
 mspI
 hpaII
 aluI
 6401 TCCGAAGTA ACTGGCTTCA GCAGAGCGCA GATACCAAT ACTGCTCTC TAGTGTAGCC CACCACITCA AGAATCTGT AGCAGCGCT
 AGGCTTCCAT TGACCGAAGT CGTCTCGCT CTATGTTTA TGACAGGAAG ATCATATCG CATCAATCG GTGGTGAAGT TCTTGAGACA TCGTGGCGGA
 mspI
 hpaII
 bsaWI
 maeIII
 hhaI/cfoI
 6501 ACATACCTCG CTCGTCTAAT CCGTGTACCA GTGGCTGCTG CCAGTGGCGA TAAGTGTGT CTTACCGGT TGGACTCAAG ACGATAGTTA CCGATAAGG
 TGTATGGAGC GAGACGATTA GGACAATGGT CACCGACGAC GGTACCGCT ATTCAGCACA GAATGGCCCA ACCTGATTC TGCTATCAAT GGCCTATTCC
 hgiAI/aspHI
 bspI286
 bsiHKA1
 bmyI
 apaLI/snoI
 alw44I/snoI
 aluI
 6601 CGCGCGGTC GGGCTGAACG GGGGTTCTGT GCACACAGCC CAGCTTGGAG CGAACGACCT ACACCGAAT GAGATACCTA CAGCGTGAGC ATTGAGAAAG
 CGGTGCGCAG CCGACTTG CCCCCAAGCA CGTGTGTGCG GTCGAACCTC GCTTGTGGA TGTGGCTTGA CTCTATGGAT GTGCACTCG TAACTCTTTC
 hinPI
 hhaI/cfoI
 haeII

SUBSTITUTE SHEET (RULE 26)

FIG. 10S

[illegible]

FIG. 10T

FIG. 10T

7101 CGTTGGCCGA TTCATTATATC CAGCTGAC
GCAACCGGCT AAGTAATTAG GTCGAC

scrFI
mvaI
ecoRII
dsav
bstNI
apyI|d
bsaJI

apyl dcm+ }	mspI	aciI	bsrBI	alul	nleIII	xmnI				
bsaJI	hpaII					asp700				
7201	CACCCCAAGC	TTTACACTTT	ATGCTTCCGG	CTCGTATGTT	GTGTGGAATT	GTGAGCGGAT	AACAATTTCACACAGGAAC	AGCTATGACC	ATGATTACGA	
	GTGGGGTCCG	AAATGTGAAA	TACGAAGGCC	GAGCATACAA	CACACCTTAA	CACTCGCCTA	TTGTTAAAGT	GTGTCTTTTG	TGATACTGG	TACTAATGCT

tru9I
mseI
aseI/asnI/vspI
7301 ATTA
TAATT

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>length: 7305
```

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/US 95/09576

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;, pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors'	1-3,7,8
Y	see page 192, line 7 - page 196, line 5; figures 2,3 --- -/--	5,6, 9-12, 16-21

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
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- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *&* document member of the same patent family

Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Hornig, H

INTERNATIONAL SEARCH REPORT

onal Application No
PCT/US 95/09576

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CELL, vol. 37, no. 3, July 1984 CELL PRESS, CAMBRIDGE, MA, US;, pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application	1-3,7,8
Y	pZIP-Neo SV(B)1 see figure 1	5,6, 9-12, 16-21
Y	--- MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC, US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1	5,6, 9-12, 16-21
Y	WO,A,94 05784 (US) 17 March 1994 see the whole document	5,6, 9-12, 16-21
Y	--- EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987 see the whole document	5,6, 9-12, 16-21
A	--- WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document	1-21
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A	--- EP,A,0 160 457 (GENENTECH INC) 6 November 1985 cited in the application see the whole document	1-21

-/--

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 95/09576

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL. ACAD. SCI., vol. 86, February 1989 NATL. ACAD SCI., WASHINGTON, DC, US;, pages 1041-1045, M. VIVAUD ET AL. 'A 5' splice-region G-C mutation in exon 1 of the human beta-globin gene inhibits pre-mRNA splicing: A mechanism for beta+-thalassemia' see the whole document -----</p>	1-4

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 95/09576

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